

CNS Spectrums

<http://journals.cambridge.org/CNS>

Additional services for **CNS Spectrums**:

Email alerts: [Click here](#)

Subscriptions: [Click here](#)

Commercial reprints: [Click here](#)

Terms of use : [Click here](#)

Mesolimbic dopamine and its neuromodulators in obesity and binge eating

Lindsay Naef, Kimberley A. Pitman and Stephanie L. Borgland

CNS Spectrums / *FirstView* Article / October 2015, pp 1 - 10

DOI: 10.1017/S1092852915000693, Published online: 30 October 2015

Link to this article: http://journals.cambridge.org/abstract_S1092852915000693

How to cite this article:

Lindsay Naef, Kimberley A. Pitman and Stephanie L. Borgland Mesolimbic dopamine and its neuromodulators in obesity and binge eating. CNS Spectrums, Available on CJO 2015 doi:10.1017/S1092852915000693

Request Permissions : [Click here](#)

Mesolimbic dopamine and its neuromodulators in obesity and binge eating

Lindsay Naef, Kimberley A. Pitman, and Stephanie L. Borgland*

Department of Physiology & Pharmacology, Cumming School of Medicine, Hotchkiss Brain Institute, University of Calgary, Calgary, Alberta, Canada

Obesity has reached epidemic prevalence, and much research has focused on homeostatic and nonhomeostatic mechanisms underlying overconsumption of food. Mesocorticolimbic circuitry, including dopamine neurons of the ventral tegmental area (VTA), is a key substrate for nonhomeostatic feeding. The goal of the present review is to compare changes in mesolimbic dopamine function in human obesity with diet-induced obesity in rodents. Additionally, we will review the literature to determine if dopamine signaling is altered with binge eating disorder in humans or binge eating modeled in rodents. Finally, we assess modulation of dopamine neurons by neuropeptides and peripheral peptidergic signals that occur with obesity or binge eating. We find that while decreased dopamine concentration is observed with obesity, there is inconsistency outside the human literature on the relationship between striatal D₂ receptor expression and obesity. Finally, few studies have explored how orexigenic or anorexigenic peptides modulate dopamine neuronal activity or striatal dopamine in obese models. However, ghrelin modulation of dopamine neurons may be an important factor for driving binge feeding in rodents.

Received 28 August 2015; Accepted 22 September 2015

Key words: Diet-induced obesity, hormones, mesolimbic dopamine, neuropeptides, striatum, ventral tegmental area.

Introduction

The current obesity epidemic has provided a strong impetus for research aimed at investigating the neurobiological basis of overeating. Mesocorticolimbic dopamine circuits are critically involved in motivated behavior. With cell bodies originating in the ventral tegmental area (VTA) and projecting to various regions including the nucleus accumbens (NAc), caudate putamen, amygdala, and prefrontal cortex,¹ dopamine neurons have long been studied in the context of motivation and drug addiction, as all drugs of abuse share the ability to stimulate dopamine neurotransmission.² Recently, dopamine circuits have emerged as an important mediator of food intake and overeating, and thus represent an ideal target for the development of drug therapies aimed at curbing overeating. The goal of the present review is to compare changes in mesolimbic dopamine function in human obesity with diet-induced obesity in rodents. Additionally, we will review the

literature to determine if dopamine signaling is altered with binge eating disorder in humans or binge eating modeled in rodents. Finally, we assess modulation of dopamine neurons by neuropeptides and peripheral peptidergic signals occurring with obesity or binge eating. Obesity and binge eating represent 2 types of overeating, and both pathologies reveal significant alterations in dopamine function. Contrasting the similarities and differences in dopamine function between the two might provide important insights into these pathophysiological states. In the *Diagnostic and Statistical Manual of Mental Disorders*, Fifth Edition (DSM-5),³ binge eating disorder (BED) is defined as “recurring episodes of eating significantly more food in a short period of time than most people would eat under similar circumstances, with episodes marked by feelings of lack of control.” Similar to humans, rodent obesity is defined as increased weight and adipose tissue compared to controls and often co-occurring with acquired insulin resistance and hyperleptinemia.⁴ In the first part of this review, we will contrast dopamine function in obesity and BED, with an emphasis on dopamine metabolism in striatal regions, the activity of the dopamine transporter, the main uptake mechanism of dopamine in the

* Address for correspondence: Stephanie Borgland, PhD, 3330 Hospital Drive Northwest, Calgary, Alberta T2N 4N1, Canada.
(Email: slborgland@ucalgary.ca)

striatum, and the dopamine D₂ receptor. In the second section, we will discuss how the hormonal and neuropeptide profiles observed in obesity and binge eating might explain the differences and similarities in dopamine function observed in the first section of the review.

Contrasting Mesolimbic Dopamine Function in Obesity and Binge Eating Disorder

Dopamine metabolism

Both human and animal studies demonstrate that striatal responses are altered in diet-induced obesity. In human studies, the direction of these effects is inconsistent. For example, functional magnetic resonance imaging (fMRI) studies using human obese subjects have demonstrated blunted striatal responses to the receipt of palatable foods^{5–7} and a cue predicting sucrose,⁸ but increased striatal responses to visual palatable food stimuli.^{9–11} Based on these divergent findings, Carnell *et al*¹² have proposed a “dynamic” model of striatal modulation in obesity with hypersensitivity to visual stimuli and hyposensitivity to the consumption of food reward. The rodent literature reveals more consistent results, with most studies showing blunted striatal dopamine in diet-induced obesity. Most studies use a 60% high fat diet or a cafeteria diet to induce obesity in rats or mice. However, there is considerable variability in the duration and onset of the exposure to the test diet. For the purposes of this review, only experiments using long-term dietary manipulations (minimum of 6 weeks) and demonstrating significant weight gain will be discussed. Diet-induced obesity is associated with decreased tyrosine hydroxylase (TH) expression, the rate-limiting factor in the synthesis of dopamine in dopamine-relevant brain regions.^{13–15} Furthermore, blunted striatal dopamine concentrations in diet-induced obesity have been described using several techniques, including total dopamine concentrations and/or dopamine turnover in the striatum in brain punches,^{16,17} decreased extracellular concentrations measured with microdialysis,^{18,19} and decreased evoked release with electrochemistry.¹⁸ Only one study revealed an increase in nucleus accumbens dopamine measured with microdialysis in obesity prone rats on an 8-week high fat diet.²⁰ Taken together, in rodent models of diet-induced obesity, there is a decrease in striatal dopamine concentration.

A limited number of studies have measured dopamine concentration in subjects with binge eating disorder. An interesting study conducted by Wang *et al*²¹ found that food stimuli and the administration of methylphenidate (a weak, long-acting catecholamine transporter inhibitor) induced a significant increase in striatal dopamine in obese binge eaters but not in obese subjects without binge eating disorder. This study suggests that dopamine

tone is elevated in binge eating disorder. Elevated striatal dopamine tone has also been observed in animal models of binge eating, including rats given intermittent access to 10% sucrose and chow followed by 12 hours of food restriction²² and in rats on a restricted feeding schedule with access to sucrose or water followed 2 hours later by chow.²³ It is important to note that in these animal studies, periods of food restriction are used to induce bingeing behavior. A large body of evidence indicates that food restriction and weight loss augment dopamine tone (reviewed in Carr²⁴), limiting our ability to make conclusions about dopamine tone enhancement in these bingeing models. While c-fos activation in VTA TH positive neurons has been observed with an intermittent access to high fat diet model of binge eating that does not require food restriction,²⁵ dopamine metabolism in animal models of bingeing without food restriction has not been measured. Taken together, preliminary evidence suggests that diet-induced obesity and binge eating may have different effects on dopamine concentration in the NAc, such that dopamine concentration is decreased in obesity, but likely increased with binge eating. Further studies are required to determine if this difference is due to a degree of adiposity associated with obesity and not binge eating, or if there are other factors underlying this difference.

D2 receptors

Most studies examining the relationship between human obesity and dopamine D₂ (and D₄) receptor availability have revealed a decrease in D₂ receptor availability in obesity,^{26–28} although increased availability has also been reported in fasted obese subjects.²⁹ One important caveat with some of the human literature is that raclopride, which is used in positron emission tomography (PET) studies, has lower affinity for the D₂ receptor than dopamine.³⁰ Thus, conditions with high dopamine concentration would appear as low D₂ receptor binding. However, other studies using fallypride have also demonstrated decreased D₂ receptor availability obese subjects.³¹ An important question is what is the underlying cause of decreased D₂ receptor availability in obesity? For example, overconsumption leading to obesity may induce decreased D₂, genetic differences in expression of D₂ receptor leading to increased susceptibility of weight gain, or both factors may occur together. Variants in the Taq1A allele of the *ANKK1* gene (neighboring the 3' untranslated region of the *DRD2* gene)³² are associated with decreased D₂ receptor expression.³³ Some studies have observed positive correlations between the Taq1A allele with body mass index,^{34,35} while others have failed to observe this correlation.³⁶ Additionally, the *DRD4-L* allele has also been associated with higher body mass index (BMI) in

humans, including individuals with bulimia nervosa.³⁷ Taken together, high BMI is associated with decreased D₂ receptor availability in the striatum. However, it is important to consider the metabolic state of the individual (fasted or sated) when they are being assessed, and whether obesity induces decreased D₂ receptor function or if this is a trait that infers susceptibility to obesity.

In rodents, studies examining the effects of diet-induced obesity on D₂ receptor expression have yielded mixed results. High-fat feeding and cafeteria feeding have been shown to reduce^{38–40} and increase^{41–43} striatal D₂ receptor expression. Putative reasons for differences include the availability of choice,⁴⁴ the macronutrient composition of the diet,⁴⁵ and the method employed to measure D₂ receptor expression. In an attempt to identify important factors, we mapped out differences in D₂ receptor expression across different studies (mostly focusing on animal studies and receptor expression, not mRNA) and identified the region of the striatum examined (ventral vs. dorsal), the species (rat vs. mouse), the type of diet, and the length of diet exposure. However, as observed in Figure 1, obesity does not predict alterations in D₂ receptor expression. One important caveat with many of these studies is that antibodies for D₂ receptors are generally not selective, and many of these studies did not show controls for D₂ antibody selectivity. Thus, more studies using more selective methods to measure D₂ receptor expression or function are needed to clarify how the development of diet-induced obesity impacts striatal D₂ receptors.

The association between the expression and function of striatal D₂ receptors and binge eating behavior has received very little attention in the literature. In a study comparing the genotypes of obese individuals with and without binge eating disorder, Davis *et al.*⁴⁶ report that

binge eating is associated with increased frequency of a gain of function allele (A2 homozygosity) in the dopamine D₂ receptor gene. Furthermore, in rodents, bingeing on sugar is associated with a significant decrease in D₂ receptor binding.^{40,47} Thus, future studies are required to assess if binge eating is associated with alterations in D₂ receptor expression or function.

The dopamine transporter (DAT)

Animal studies have reported alterations in DAT expression and function in diet-induced obesity, although human studies have yet to reveal consistent effects. Thomsen *et al.*⁴⁸ observed no correlation between body mass index and striatal DAT availability in the striatum, caudate nucleus, and putamen and no significant group difference between obese and severely obese and normal weight controls. Similarly, in a study of 123 participants, the authors report no association between BMI and striatal DAT availability.⁴⁹ However, low striatal DAT levels were associated with elevated BMI in 50 human participants using single-photon emission computed tomography (SPECT).⁵⁰ Mixed results were obtained in animal models of obesity. Cone *et al.*⁵¹ report a deficit in the rate of dopamine uptake by the DAT in rats exposed to a high-fat diet for 6 weeks but no change in mRNA. It is important to note that the rats did not develop obesity in this study. Likewise, South *et al.*⁴² and Narayanaswami *et al.*²⁰ report a reduction in DAT expression in obese rodents. Importantly, many of these studies use a variety of types and duration of diets to induce obesity, and a few measure hyperleptinemia or insulin resistance. Given that both leptin and insulin can also modulate DAT expression and function,^{52,53} it is important to consider the metabolic state of the animal in these experiments.

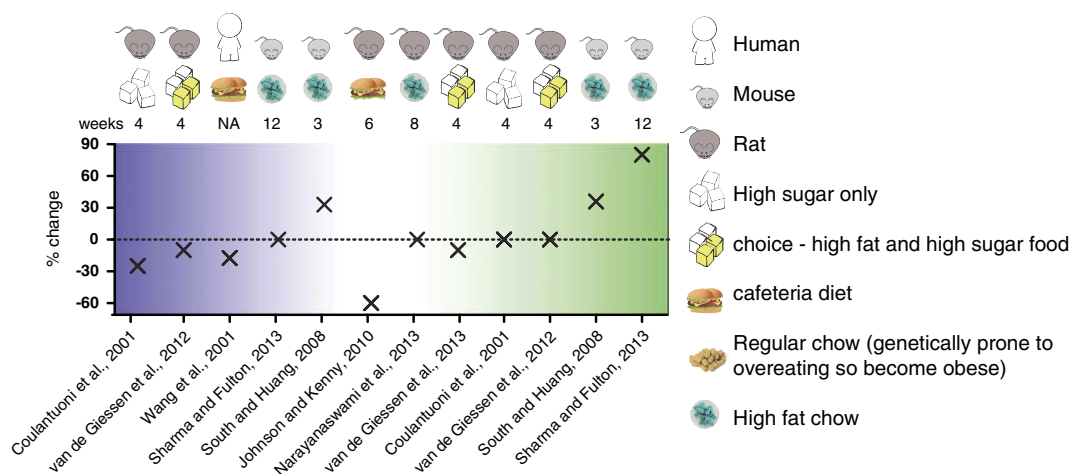


FIGURE 1. Dopamine D₂ expression in diet-induced obesity. Variations in the effects of diet composition and duration on D₂ receptors expression (% of experimental control group in each study) in subregions of the striatum (purple: dorsal striatum, green: ventral striatum, white: unspecified). Some studies report more than one subregion and are thus, represented for each subregion.

Furthermore, a recent study by Hryhorczuk *et al*⁵⁴ demonstrates that the type of dietary fat is an important determinant of DAT function, as exposure to saturated fat, but not monounsaturated fat, induced significant reductions in DAT expression. Taken together, in rodents, obesity, leptin, insulin, and saturated fat can modulate DAT number or function. In human studies, there is no consistent effect of DAT availability in obese subjects.

The relationship between binge eating disorder and DAT has exclusively been examined in studies aimed at identifying genetic polymorphisms in dopamine transporter genes. To date, one polymorphism for DAT has been identified. In the 3' untranslated region of the dopamine transporter gene (*DAT1*), the frequency of a short allele was significantly higher in a cohort of Japanese women with BED, and this polymorphism has been associated with lower DAT function.⁵⁵ However, a Canadian study failed to replicate these findings.⁵⁶ In an animal model of bingeing, restricted feeding with scheduled sucrose access resulted in an upregulation of the rat DAT.⁵⁷ Taken together, binge eating may be associated with increased dopamine reuptake.

Conclusions

Deficits in dopamine function observed in humans and animals have led to the “reward deficiency” hypothesis of obesity,⁵⁸ which proposes that reduced dopamine tone leads to overeating as an attempt to restore striatal dopamine concentrations. However, it is important to also consider dopamine’s role in energy expenditure, such that decreased dopamine function may simply reduce movement and thus energy expenditure, leading to obesity.⁵⁹ Furthermore, striatal dopamine is thought to encode a reward prediction error (the difference between actual and expected reward), therefore reduced blood-oxygen-level dependent (BOLD) signal or dopamine concentration upon food receipt may result from increased reward anticipation and hence a reduced reward prediction error signal.

Influence of Obesity and Binge Eating on Orexigenic or Anorexigenic Peptide Modulation of Dopamine Neurons

A variety of neuropeptides and circulating hormones acts on VTA dopamine neurons to influence their activity and output. For example, peptides that typically promote food intake increase the activity of dopamine neurons and dopaminergic output. In contrast, anorexigenic peptides administered in the VTA decrease food consumption, but their modulation of dopamine does not necessarily predict feeding. Most studies examining the role of neuropeptides and peripheral hormones on

dopamine neurons have been explored in naïve animals, and this work has been reviewed in detail elsewhere.⁶⁰ However, less is known about how dopamine neurons are modulated by peptides during obesity or during binge eating. In this section, we outline the evidence for orexigenic and anorexigenic modulators of dopamine neurons in obesity and binge eating.

Ghrelin

Ghrelin, a peptide hormone produced from the enteroendocrine cells of the stomach,⁶¹ is implicated in hunger and meal initiation. Circulating levels increase prior to expected mealtimes and decrease with feeding.⁶² Ghrelin injections robustly stimulate food intake rapidly and transiently primarily by increasing appetitive feeding behaviors.⁶³ In naïve adult animals, ghrelin in the VTA increases locomotor activity, palatable food consumption, and a robust motivated feeding response via its only known receptor, growth hormone secretagogue receptor (GHSR).^{63–67} Furthermore, ghrelin increases excitatory plasticity onto dopamine neurons⁶⁸ and increases dopamine release in the NAc.^{51,63,65,69,70} Therefore, ghrelin’s actions at promoting appetitive behaviors may be mediated by the VTA.

In opposition to increased ghrelin levels in response to energy deficiency, ghrelin levels are depressed with weight gain resulting from high caloric diets^{71–73} or overfeeding by intragastric gavage.⁷⁴ Because ghrelin action on dopamine neurons increases food motivation, one might expect alterations in ghrelin modulation of the mesolimbic system in obesity. Indeed, it has been proposed that ghrelin’s ability to enhance the reward value of food by its actions in the VTA might drive overeating, leading to obesity.^{75,76} However, it is not clear how ghrelin signals in the VTA once obesity has set in. Using search terms such as “ghrelin/VTA/obese” or “ghrelin/dopamine/obese” in PubMed, it appears that there are no published reports of alterations of ghrelin signaling in the VTA or ghrelin-mediated dopamine output using models of obese animals. In a rodent model of binge eating disorder, where *ad libitum* fed rodents were exposed to a high fat diet 2 hours per day for 4 days (intermittent access to a high-fat diet), significant c-fos activation in VTA dopamine neurons was observed with escalating food intake.²⁵ This effect was not observed in GHSR knockout mice or in high-fat diet fed controls,²⁵ which suggests that ghrelin signaling is required for escalation of food intake associated with binge eating.

Orexin

Intracranial administration of the lateral hypothalamic peptide orexin, also known as hypocretin, promotes

food intake.⁷⁷ Interestingly, ghrelin-induced conditioned place preference for high-fat food⁷⁸ or ghrelin-induced food intake⁷⁹ was blocked with orexin receptor antagonists in the VTA, suggesting that ghrelin modulation of reward value of food requires orexin signaling. Importantly, orexin modulates VTA dopamine neurons to promote motivated food intake.⁸⁰ Orexin increases neuronal firing,⁸¹ synaptic efficacy,⁸² and output^{83,84} of dopamine neurons in naïve animals. In rats that have been self-administering high-fat pellets, orexin-mediated potentiation of excitatory synaptic transmission onto dopamine neurons is enhanced.⁸⁵

Using obesity models, reports have indicated that there is increased expression of orexin 1 (OX1R) and orexin 2 (OX2R) receptors in the rostral lateral hypothalamus of obesity prone, but not obesity resistant, Sprague Dawley rats on a chow diet.⁸⁶ Notably, the obesity resistant rats had increased food consumption and spontaneous locomotor activity induced by intracerebroventricular orexin administration,⁸⁶ suggesting that orexin signaling may provide resistance to the development of obesity. Consistent with this, mice overexpressing orexin peptides on a high-fat diet were resistant to weight gain compared to wild type mice, due to increased energy expenditure. These effects were mediated via the OX2R.⁸⁷ Mice already obese from an 8-week high-fat diet showed decreased number of immunopositive orexin neurons in the lateral hypothalamus,⁸⁸ which might decrease orexin's impact energy expenditure once animals are obese. In an obese state, where there is hyperleptinemia and leptin receptor deficiency, one might predict an increase in orexin-induced feeding, as leptin is known to suppress this activity in naïve animals.^{89,90} However, leptin and orexin appear to act cooperatively to coordinate energy sensing and behavior. In naïve animals, orexin neurons receive primarily excitatory input expressing cannabinoid 1 receptors (CB1Rs). However, in the obese state, the ratio of CB1-expressing inputs to orexin neurons is predominantly inhibitory. In obese animals, leptin treatment restores excitatory CB1R-terminal bias onto orexin neurons similar to that of lean mice.⁹¹ Taken together, obesity changes signaling at inputs onto orexin neurons that may influence orexin release in target regions.

In the VTA, a 6-week high-fat diet increases OX1R expression compared to chow-fed controls.⁹² However, so far, it remains to be determined how orexin signaling in the VTA is altered in the obese state. Acute high fat diet consumption (2 h) increases c-fos activation of both orexin neurons and that of dopamine neurons in the VTA. Increased high-fat diet-induced c-fos activation in the VTA is blocked by a systemic OX1R antagonist.⁹³ Using a binge-feeding model of intermittent access to a high-fat diet, these authors found increased c-fos expression in the VTA and escalation of food

consumption. However, this was not blocked with an OX1R antagonist.²⁵ Taken together, these studies suggest that while orexin signaling may be protective against obesity, orexin signaling may be altered once animals are in an obese state. A high-fat diet may increase activation of dopamine neurons in an orexin-dependent manner, but orexin signaling does not appear to underlie escalation of food intake associated with binge eating.

Anorexigenic peptides

Leptin

Hyperleptinemia and leptin receptor (LepRb) resistance is typically observed in the arcuate nucleus during obesity.^{94,95} In addition to targeting hypothalamic circuits, leptin signals on VTA dopamine neurons via activation of LepRb and phosphorylation of signal transducer and activator of transcription 3 (pSTAT3)^{39,40} or extracellular signal-regulated kinase-1 and -2 (pERK1/2).⁹⁶ Leptin decreases the firing rate of VTA dopamine neurons^{96,97} and suppresses excitatory synaptic transmission onto dopamine neurons.⁹⁸ Leptin's effects on dopamine release are dependent on the satiety and motivational state of the animal.⁶⁰ For example, in food-restricted animals, leptin decreases dopamine release in the NAc⁹⁹ and food-cue-induced dopamine release.¹⁰⁰ In leptin deficient ob/ob mice, evoked dopamine release in the NAc is diminished, but can be restored with leptin administration into the VTA or NAc.^{101,102} In mice fed a high fat diet for 6 months or in mice overexpressing leptin, cellular resistance to leptin signaling was observed in the VTA and the arcuate nucleus, but not in other hypothalamic regions.¹⁰³ Furthermore, central administration of leptin diminished preference for palatable food in the control group, but not in the obese animals.¹⁰³ In contrast, intra-VTA leptin decreased food intake in Sprague Dawley rats on a 16-week high fat diet similar to rats on a control diet.¹⁰⁴ However, there was no difference in weight gain on these diets, nor was peripheral leptin resistance measured, and therefore, it was unclear if hyperleptinemia occurred. When the high-fat diet-fed rats were separated into top and bottom quartiles of weight gain, those in the bottom quartile (diet-resistant) had a greater intra-VTA leptin-induced inhibition of food intake compared to rats in the top quartile (diet-induced obesity), indicating the possibility of leptin resistance in the VTA of rats that gain the most weight.¹⁰⁴ In contrast to this, male Wistar rats were given free choice to a high-fat, high-sugar diet for 7 days that induced peripheral hyperleptinemia, but no change in central leptin's effect on food intake.¹⁰⁵ Central leptin decreased TH expression in the VTA of control rats, but not those fed the high fat/high sugar diet for 7 days. Taken together, the duration and type of diet may play a key role in determining if leptin

resistance can occur in the VTA, and if this may influence leptin's effect on food intake.

Insulin

Plasma insulin levels rise prior to meal consumption in a cephalic response to cues predicting food, including sight, smell, and mealtime.¹⁰⁶ Higher concentrations of insulin are released postprandially and can act in the ventral medial hypothalamus and VTA to inhibit food intake.¹⁰⁷ Insulin receptors as well as intracellular substrates of insulin receptor activation, including insulin receptor substrate 2 (IRS2) and phosphatidylinositol (3,4,5)-triphosphate, a product of phosphatidylinositol 3 kinase (PI3K), are expressed on dopamine neurons.^{108–110} Insulin in the VTA induces a long-term depression (LTD) that is mediated by endocannabinoid suppression of excitatory synapses.¹¹¹ This LTD is selective for excitatory but not inhibitory synapses onto dopamine neurons of the VTA.¹¹¹ Insulin in the VTA also decreases somatodendritic dopamine via a PI3K dependent mechanisms and increased reuptake through dopamine transporters.⁵³ In normal weight animals, insulin in the VTA decreases hedonic feeding in sated animals⁵³ or that evoked by administration of mu-opioids to the VTA.⁴⁹ Consistent with this effect, insulin in the VTA can increase the threshold for brain stimulation reward.¹⁰⁸ Finally, intra-VTA insulin decreases preference for a context previously associated with palatable food in a dose dependent manner and reduces food anticipatory behaviors, but has no effect on the effort required to obtain palatable reinforcers.¹¹² Few studies have explored how intra-VTA insulin signaling or insulin's effects on food intake are altered by obesity. However, in mice with chronic loss of insulin receptors on TH-positive neurons exhibit increased body weight, fat mass, and hyperphagia.¹¹³ Furthermore, in a mouse strain exhibiting higher plasma insulin levels, insulin-induced LTD onto dopamine neurons was suppressed, even though other forms of synaptic transmission onto dopamine neurons were intact.¹¹⁰ In a model of early life obesity, whereby litter 'size was restricted, postnatal overfeeding led to an increase in insulin's ability to induce phosphorylation of downstream signaling cascades in the VTA, indicating increased insulin receptor sensitivity on dopamine neurons without any changes in peripheral insulin signaling.¹¹⁴ These results suggest the possibility that insulin signaling in the VTA may be a region that does not succumb to insulin resistance, as has been observed with some areas in the hypothalamus.¹¹⁵ The relevance of regional differences in obesity-induced insulin receptor insensitivity has not been determined. However, one can speculate that some intracellular cascades coupling to insulin receptors may be more vulnerable to desensitization than others. Taken together, insulin in the VTA decreases excitatory transmission to dopamine neurons as

well as different aspects of hedonic food intake. Further study is required to determine if insulin resistance in the VTA can occur with obesity.

Glucagon-like peptide (GLP-1)

GLP-1 is released from the distal gut and neurons of the nucleus tractus solitarius (NTS) and stimulates insulin release from pancreatic beta-cells.^{116–118} GLP-1 neurons of the NTS make monosynaptic connections with neurons of the VTA and NAc,¹¹⁹ and GLP-1 receptors are expressed in both of these regions.^{117,120} Intra-VTA application of the stable GLP-1 agonist, exendin-4, increases TH expression and decreases food intake, possibly via modestly increasing presynaptic glutamate release.¹²¹ Consistent with this report, chemogenetic-mediated endogenous release of GLP-1 from NTS terminals in the VTA decreased high-fat food intake, an effect that was blocked by a systemic GLP-1 antagonist, exendin-9.¹²² However, in contrast to previous studies, stimulation of GLP-1 receptors on NAc-projecting dopamine neurons suppressed excitatory, but not inhibitory, synaptic transmission.¹²² This discrepancy is likely due to differences in GLP-1 receptor expressing subpopulations of dopamine neurons. Because GLP-1 plays a role in stimulating insulin release, it has been proposed to be an anti-obesity agent.¹²³ Indeed, chronic exendin-4 in weaned rats¹²⁴ and short-term exendin-4 in pre-weaned rats¹²⁵ reverse effects of diet-induced obesity and insulin resistance in offspring from obese dams. Interestingly, a systemic GLP-1R agonist can reduce binge eating in female mice on an intermittent high-fat diet.¹²⁶ At the time of writing, there are no reports on whether GLP-1R agonists administered in the VTA can reduce feeding in obese or binge-eating animals. However, a GLP-1R agonist has shown to be effective in reducing weight gain, cardiovascular measures, and binge eating episodes in obese nondiabetic humans,¹²⁷ suggesting that GLP-1 receptors may be a promising pharmacotherapeutic target for treatment of binge eating.

Conclusion and Future Considerations

In obesity, human studies reveal decreased activation of regions involved in food reward upon the presentation of food-predicting cues and upon consumption of palatable foods. Consistent with this, most rodent models of diet-induced obesity show reduced striatal dopamine concentration due to decreased TH expression or increased dopamine reuptake. Decreased D₂ receptor expression is associated with increased BMI in humans. However, it is important to consider that different radioligands used for PET imaging may give different results as well as the metabolic state of the individual (ie if the individual

is fasted) may influence D₂ receptor occupancy. In rodent studies, there appears to be no consensus on whether D₂ receptor expression is altered with obesity. However, there are several issues that may influence the differing results among these studies. First, the type (high-fat, high-sucrose, or cafeteria diet) and duration (anywhere from 5 days to 16 weeks) of the diet may influence D₂ receptor expression. Second, the strategy used to measure D₂ receptor expression may also influence the findings.

Neuromodulation of VTA dopamine neurons by orexigenic or anorexigenic peptides in obese animals has only begun to be explored. While orexin signaling may be protective against obesity due its effect on increased energy expenditure,⁸⁶ it is unclear if any of these effects is mediated by the VTA. Obesity increases inhibitory input to lateral hypothalamic orexin neurons⁹¹; thus reduced orexin release in target regions would likely be expected with obesity. Consistent with this, increased OX1Rs were observed in the VTA with obesity.⁹² Anorexigenic agents such as insulin or leptin also target the VTA. In diet-induced obesity, studies conflict as to whether leptin or insulin signaling becomes desensitized. This factor may be due to the types and durations of diets used, and thus, direct measures of peripheral leptin or insulin resistance should be recorded along with measures of leptin or insulin signaling in the VTA. Indeed regionally selective insulin resistance has been observed in the hypothalamus, with the arcuate nucleus most susceptible to insulin resistance.¹¹⁵

Binge eating does not necessarily require animals or humans to be obese, and therefore different alteration in dopamine signaling may be expected. Indeed, dopamine metabolism may be increased in binge eating humans. In rodents, food restriction is often employed to promote escalating food intake modeling binge eating. However, because food restriction and weight loss can augment dopamine tone,²⁴ it is difficult to draw conclusions on how dopamine is modulated during binge eating. However, increased activation of VTA dopamine neurons was observed using intermittent access to a high-fat diet, which promotes escalation of food intake without food restriction.²⁵ This effect was dependent on ghrelin, but not orexin, signaling.²⁵ Future studies are required to elucidate the mechanism associated with increased c-fos expression in binge eating animals. Finally, targeting insulin, leptin, or GLP-1 receptor, which are known to depress synaptic transmission onto dopamine neurons, may provide a good pharmacotherapeutic strategy for treatment of binge eating disorders.

Disclosures

Stephanie Borgland has the following disclosures: Canadian Institutes of Health Research, Principal Investigator, CIHR

Grant MOP 102617; Natural Science and Engineering Research Council, Principal Investigator, NSERC Grant MOP 372517. Kimberley Pitman and Lindsay Naef have nothing to disclose.

REFERENCES:

1. Beier KT, Steinberg EE, DeLoach KE, *et al.* Circuit architecture of VTA dopamine neurons revealed by systematic input-output mapping. *Cell*. 2015; **162**(3): 622–634.
2. Di Chiara G, Imperato A. Drugs abused by humans preferentially increase synaptic dopamine concentrations in the mesolimbic system of freely moving rats. *Proc Natl Acad Sci U S A*. 1988; **85**(14): 5274–5278.
3. Call C, Walsh BT, Attia E. From DSM-IV to DSM-5: changes to eating disorder diagnoses. *Curr Opin Psychiatry*. 2013; **26**(6): 532–536.
4. Woods SC, Seeley RJ, Rushing PA, D'Alessio D, Tso P. A controlled high-fat diet induces an obese syndrome in rats. *J Nutr*. 2003; **133**(4): 1081–1087.
5. Stice E, Spoor S, Bohon C, Small DM. Relation between obesity and blunted striatal response to food is moderated by TaqIA A1 allele. *Science*. 2008; **322**(5900): 449–452.
6. Stice E, Spoor S, Bohon C, Veldhuizen MG, Small DM. Relation of reward from food intake and anticipated food intake to obesity: a functional magnetic resonance imaging study. *J Abnorm Psychol*. 2008; **117**(4): 924–935.
7. Green E, Jacobson A, Haase L, Murphy C. Reduced nucleus accumbens and caudate nucleus activation to a pleasant taste is associated with obesity in older adults. *Brain Res*. 2011; **1386**: 109–117.
8. Frank GKW, Reynolds JR, Shott ME, *et al.* Anorexia nervosa and obesity are associated with opposite brain reward response. *Neuropsychopharmacology*. 2012; **37**(9): 2031–2046.
9. Martin LE, Holsen LM, Chambers RJ, *et al.* Neural mechanisms associated with food motivation in obese and healthy weight adults. *Obesity (Silver Spring)*. 2010; **18**(2): 254–260.
10. Rothemund Y, Preuschhof C, Böhner G, *et al.* Differential activation of the dorsal striatum by high-calorie visual food stimuli in obese individuals. *Neuroimage*. 2007; **37**(2): 410–421.
11. Stoeckel LE, Kim J, Weller RE, Cox JE, Cook EW, Horwitz B. Effective connectivity of a reward network in obese women. *Brain Res Bull*. 2009; **79**(6): 388–395.
12. Carnell S, Gibson C, Benson L, Ochner CN, Geliebter A. Neuroimaging and obesity: current knowledge and future directions. *Obes Rev*. 2012; **13**(1): 43–56.
13. Li Y, South T, Han M, Chen J, Wang R, Huang X-F. High-fat diet decreases tyrosine hydroxylase mRNA expression irrespective of obesity susceptibility in mice. *Brain Res*. 2009; **1268**: 181–189.
14. Ong ZY, Wanasuria AF, Lin MZP, Hiscock J, Muhlhauser BS. Chronic intake of a cafeteria diet and subsequent abstinence: sex-specific effects on gene expression in the mesolimbic reward system. *Appetite*. 2013; **65**: 189–199.
15. Ahmed S, Kashem MA, Sarker R, Ahmed EU, Hargreaves GA, McGregor IS. Neuroadaptations in the striatal proteome of the rat following prolonged excessive sucrose intake. *Neurochem Res*. 2014; **39**(5): 815–824.
16. Davis JF, Tracy AL, Schurdak JD, *et al.* Exposure to elevated levels of dietary fat attenuates psychostimulant reward and mesolimbic dopamine turnover in the rat. *Behav Neurosci*. 2008; **122**(6): 1257–1263.
17. Zhang C, Wei N-L, Wang Y, Wang X, Zhang J-G, Zhang K. Deep brain stimulation of the nucleus accumbens shell induces anti-obesity effects in obese rats with alteration of dopamine neurotransmission. *Neurosci Lett*. 2015; **589**: 1–6.

18. Hansen HH, Jensen MM, Overgaard A, Weikop P, Mikkelsen JD. Tesofensine induces appetite suppression and weight loss with reversal of low forebrain dopamine levels in the diet-induced obese rat. *Pharmacol Biochem Behav.* 2013; **110**: 265–271.
19. Geiger BM, Haburcak M, Avena NM, Moyer MC, Hoebel BG, Pothos EN. Deficits of mesolimbic dopamine neurotransmission in rat dietary obesity. *Neuroscience.* 2009; **159**(4): 1193–1199.
20. Narayanaswami V, Thompson AC, Cassis LA, Bardo MT, Dwoskin LP. Diet-induced obesity: dopamine transporter function, impulsivity and motivation. *Int J Obes (Lond).* 2013; **37**(8): 1095–1103.
21. Wang G-J, Geliebter A, Volkow ND, *et al.* Enhanced striatal dopamine release during food stimulation in binge eating disorder. *Obesity (Silver Spring).* 2011; **19**(8): 1601–1608.
22. Rada P, Avena NM, Hoebel BG. Daily bingeing on sugar repeatedly releases dopamine in the accumbens shell. *Neuroscience.* 2005; **134**(3): 737–744.
23. Hajnal A, Norgren R. Repeated access to sucrose augments dopamine turnover in the nucleus accumbens. *Neuroreport.* 2002; **13**(17): 2213–2216.
24. Carr KD. Chronic food restriction: enhancing effects on drug reward and striatal cell signaling. *Physiol Behav.* 2007; **91**(5): 459–472.
25. Valdivia S, Cornejo MP, Reynaldo M, De Francesco PN, Perello M. Escalation in high fat intake in a binge eating model differentially engages dopamine neurons of the ventral tegmental area and requires ghrelin signaling. *Psychoneuroendocrinology.* 2015; **60**: 206–216.
26. de Weijer BA, van de Giessen E, van Amelsvoort TA, *et al.* Lower striatal dopamine D2/3 receptor availability in obese compared with non-obese subjects. *EJNMMI Res.* 2011; **1**(1): 37.
27. Haltia LT, Rinne JO, Merisaari H, *et al.* Effects of intravenous glucose on dopaminergic function in the human brain in vivo. *Synapse.* 2007; **61**(9): 748–756.
28. Wang GJ, Volkow ND, Logan J, *et al.* Brain dopamine and obesity. *Lancet.* 2001; **357**(9253): 354–357.
29. Dunn JP, Kessler RM, Feurer ID, *et al.* Relationship of dopamine type 2 receptor binding potential with fasting neuroendocrine hormones and insulin sensitivity in human obesity. *Diabetes Care.* 2012; **35**(5): 1105–1111.
30. Kung HF, Pan S, Kung MP, *et al.* In vitro and in vivo evaluation of [¹²³I]IBZM: a potential CNS D-2 dopamine receptor imaging agent. *J Nucl Med.* 1989; **30**(1): 88–92.
31. Guo J, Simmons WK, Herscovitch P, Martin A, Hall KD. Striatal dopamine D2-like receptor correlation patterns with human obesity and opportunistic eating behavior. *Mol Psychiatry.* 2014; **19**(10): 1078–1084.
32. Fossella J, Green AE, Fan J. Evaluation of a structural polymorphism in the ankyrin repeat and kinase domain containing 1 (ANKK1) gene and the activation of executive attention networks. *Cogn Affect Behav Neurosci.* 2006; **6**(1): 71–78.
33. Ritchie T, Noble EP. Association of seven polymorphisms of the D2 dopamine receptor gene with brain receptor-binding characteristics. *Neurochem Res.* 2003; **28**(1): 73–82.
34. Roth CL, Hinney A, Schur EA, Elfers CT, Reinehr T. Association analyses for dopamine receptor gene polymorphisms and weight status in a longitudinal analysis in obese children before and after lifestyle intervention. *BMC Pediatr.* 2013; **13**: 197.
35. Thomas GN, Critchley JA, Tomlinson B, Cockram CS, Chan JC. Relationships between the taq1 polymorphism of the dopamine D2 receptor and blood pressure in hyperglycaemic and normoglycaemic Chinese subjects. *Clin Endocrinol (Oxf).* 2001; **55**(5): 605–611.
36. Southon A, Walder K, Sanigorski AM, *et al.* The Taq IA and Ser311 Cys polymorphisms in the dopamine D2 receptor gene and obesity. *Diabetes Nutr Metab.* 2003; **16**(1): 72–76.
37. Kaplan AS, Levitan RD, Yilmaz Z, Davis C, Tharmalingam S, Kennedy JL. A DRD4/BDNF gene-gene interaction associated with maximum BMI in women with bulimia nervosa. *Int J Eat Disord.* 2008; **41**(1): 22–28.
38. Johnson PM, Kenny PJ. Dopamine D2 receptors in addiction-like reward dysfunction and compulsive eating in obese rats. *Nat Neurosci.* 2010; **13**(5): 635–641.
39. Hajnal A, Margas WM, Covasa M. Altered dopamine D2 receptor function and binding in obese OLETF rat. *Brain Res Bull.* 2008; **75**(1): 70–76.
40. Colantuoni C, Schwenker J, McCarthy J, *et al.* Excessive sugar intake alters binding to dopamine and mu-opioid receptors in the brain. *Neuroreport.* 2001; **12**(16): 3549–3552.
41. Huang X-F, Yu Y, Zavitsanou K, Han M, Storlien L. Differential expression of dopamine D2 and D4 receptor and tyrosine hydroxylase mRNA in mice prone, or resistant, to chronic high-fat diet-induced obesity. *Brain Res Mol Brain Res.* 2005; **135**(1–2): 150–161.
42. South T, Huang X-F. High-fat diet exposure increases dopamine D2 receptor and decreases dopamine transporter receptor binding density in the nucleus accumbens and caudate putamen of mice. *Neurochem Res.* 2008; **33**(3): 598–605.
43. Sharma S, Fulton S. Diet-induced obesity promotes depressive-like behaviour that is associated with neural adaptations in brain reward circuitry. *Int J Obes (Lond).* 2013; **37**(3): 382–389.
44. van de Giessen E, la Fleur SE, de Bruin K, van den Brink W, Booij J. Free-choice and no-choice high-fat diets affect striatal dopamine D2/3 receptor availability, caloric intake, and adiposity. *Obesity (Silver Spring).* 2012; **20**(8): 1738–1740.
45. van de Giessen E, la Fleur SE, Eggels L, de Bruin K, van den Brink W, Booij J. High fat/carbohydrate ratio but not total energy intake induces lower striatal dopamine D2/3 receptor availability in diet-induced obesity. *Int J Obes (Lond).* 2013; **37**(5): 754–757.
46. Davis CA, Levitan RD, Reid C, *et al.* Dopamine for “wanting” and opioids for “liking”: a comparison of obese adults with and without binge eating. *Obesity (Silver Spring).* 2009; **17**(6): 1220–1225.
47. Bello NT, Lucas LR, Hajnal A. Repeated sucrose access influences dopamine D2 receptor density in the striatum. *Neuroreport.* 2002; **13**(12): 1575–1578.
48. Thomsen G, Ziebell M, Jensen PS, da Cuhna-Bang S, Knudsen GM, Pinborg LH. No correlation between body mass index and striatal dopamine transporter availability in healthy volunteers using SPECT and [¹²³I]PE2I. *Obesity (Silver Spring).* 2013; **21**(9): 1803–1806.
49. van de Giessen E, Hesse S, Caan MWA, *et al.* No association between striatal dopamine transporter binding and body mass index: a multi-center European study in healthy volunteers. *Neuroimage.* 2013; **64**: 61–67.
50. Chen PS, Yang YK, Yeh TL, *et al.* Correlation between body mass index and striatal dopamine transporter availability in healthy volunteers—a SPECT study. *Neuroimage.* 2008; **40**(1): 275–279.
51. Cone JJ, Chartoff EH, Potter DN, Ebner SR, Roitman MF. Prolonged high fat diet reduces dopamine reuptake without altering DAT gene expression. *PLoS One.* 2013; **8**(3): e58251.
52. Perry ML, Leininger GM, Chen R, *et al.* Leptin promotes dopamine transporter and tyrosine hydroxylase activity in the nucleus accumbens of Sprague-Dawley rats. *J Neurochem.* 2010; **114**(3): 666–674.
53. Mebel DM, Wong JCY, Dong YJ, Borgland SL. Insulin in the ventral tegmental area reduces hedonic feeding and suppresses dopamine concentration via increased reuptake. *Eur J Neurosci.* 2012; **36**(3): 2336–2346.
54. Hryhorczuk C, Florea M, Rodaros D, *et al.* Dampened mesolimbic dopamine function and signaling by saturated but not monounsaturated dietary lipids. *Neuropsychopharmacology.* In press. DOI: 10.1038/npp.2015.207.

55. Shinohara M, Mizushima H, Hirano M, *et al.* Eating disorders with binge-eating behaviour are associated with the s allele of the 3'-UTR VNTR polymorphism of the dopamine transporter gene. *J Psychiatry Neurosci.* 2004; **29**(2): 134-137.
56. Davis C, Levitan RD, Kaplan AS, *et al.* Dopamine transporter gene (DAT1) associated with appetite suppression to methylphenidate in a case-control study of binge eating disorder. *Neuropsychopharmacology.* 2007; **32**(10): 2199-2206.
57. Bello NT, Sweigart KL, Lakoski JM, Norgren R, Hajnal A. Restricted feeding with scheduled sucrose access results in an upregulation of the rat dopamine transporter. *Am J Physiol Regul Integr Comp Physiol.* 2003; **284**(5): R1260-R1268.
58. Blum K, Sheridan PJ, Wood RC, *et al.* The D2 dopamine receptor gene as a determinant of reward deficiency syndrome. *J R Soc Med.* 1996; **89**(7): 396-400.
59. Beeler JA, Faust RP, Turkson S, Ye H, Zhuang X. Low dopamine D2 receptor increases vulnerability to obesity via reduced physical activity not increased appetitive motivation. *Biol Psychiatry.* In press. DOI: 10.1016/j.biopsych.2015.07.009.
60. Liu S, Borgland SL. Regulation of the mesolimbic dopamine circuit by feeding peptides. *Neuroscience.* 2015; **289**: 19-42.
61. Kojima M, Hosoda H, Date Y, Nakazato M, Matsuo H, Kangawa K. Ghrelin is a growth-hormone-releasing acylated peptide from stomach. *Nature.* 1999; **402**(6762): 656-660.
62. Cummings DE. Ghrelin and the short- and long-term regulation of appetite and body weight. *Physiol Behav.* 2006; **89**(1): 71-84.
63. Jerlhag E, Eggecioglu E, Dickson SL, Douhan A, Svensson L, Engel JA. Ghrelin administration into tegmental areas stimulates locomotor activity and increases extracellular concentration of dopamine in the nucleus accumbens. *Addict Biol.* 2007; **12**(1): 6-16.
64. Naleid AM, Grace MK, Cummings DE, Levine AS. Ghrelin induces feeding in the mesolimbic reward pathway between the ventral tegmental area and the nucleus accumbens. *Peptides.* 2005; **26**(11): 2274-2279.
65. Kawahara Y, Kawahara H, Kaneko F, *et al.* Peripherally administered ghrelin induces bimodal effects on the mesolimbic dopamine system depending on food-consumptive states. *Neuroscience.* 2009; **161**(3): 855-864.
66. Skibicka KP, Hansson C, Alvarez-Crespo M, Friberg PA, Dickson SL. Ghrelin directly targets the ventral tegmental area to increase food motivation. *Neuroscience.* 2011; **180**: 129-137.
67. Eggecioglu E, Jerlhag E, Salomé N, *et al.* Ghrelin increases intake of rewarding food in rodents. *Addict Biol.* 2010; **15**(3): 304-311.
68. Abizaid A, Liu Z-W, Andrews ZB, *et al.* Ghrelin modulates the activity and synaptic input organization of midbrain dopamine neurons while promoting appetite. *J Clin Invest.* 2006; **116**(12): 3229-3239.
69. Quarta D, Di Francesco C, Melotto S, Mangiarini L, Heidbreder C, Hedou G. Systemic administration of ghrelin increases extracellular dopamine in the shell but not the core subdivision of the nucleus accumbens. *Neurochem Int.* 2009; **54**(2): 89-94.
70. Cone JJ, Roitman JD, Roitman MF. Ghrelin regulates phasic dopamine and nucleus accumbens signaling evoked by food-predictive stimuli. *J Neurochem.* 2015; **133**(6): 844-856.
71. Moesgaard SG, Ahrén B, Carr RD, Gram DX, Brand CL, Sundler F. Effects of high-fat feeding and fasting on ghrelin expression in the mouse stomach. *Regul Pept.* 2004; **120**(1-3): 261-267.
72. Perreault M, Istrate N, Wang L, Nichols AJ, Tozzo E, Stricker-Krongrad A. Resistance to the orexigenic effect of ghrelin in dietary-induced obesity in mice: reversal upon weight loss. *Int J Obes (Lond).* 2004; **28**(7): 879-885.
73. Lindqvist A, de la Cour CD, Stegmark A, Håkanson R, Erlanson-Albertsson C. Overeating of palatable food is associated with blunted leptin and ghrelin responses. *Regul Pept.* 2005; **130**(3): 123-132.
74. Williams DL, Grill HJ, Cummings DE, Kaplan JM. Overfeeding-induced weight gain suppresses plasma ghrelin levels in rats. *J Endocrinol Invest.* 2006; **29**(10): 863-868.
75. Murray S, Tulloch A, Gold MS, Avena NM. Hormonal and neural mechanisms of food reward, eating behaviour and obesity. *Nat Rev Endocrinol.* 2014; **10**(9): 540-552.
76. van Zessen R, van der Plasse G, Adan RA. Contribution of the mesolimbic dopamine system in mediating the effects of leptin and ghrelin on feeding. *Proc Nutr Soc.* 2012; **71**(4): 435-445.
77. Mahler SV, Moorman DE, Smith RJ, James MH, Aston-Jones G. Motivational activation: a unifying hypothesis of orexin/hypocretin function. *Nat Neurosci.* 2014; **17**(10): 1298-1303.
78. Perello M, Sakata I, Birnbaum S, *et al.* Ghrelin increases the rewarding value of high-fat diet in an orexin-dependent manner. *Biol Psychiatry.* 2010; **67**(9): 880-886.
79. Cone JJ, McCutcheon JE, Roitman MF. Ghrelin acts as an interface between physiological state and phasic dopamine signaling. *J Neurosci.* 2014; **34**(14): 4905-4913.
80. Thompson JL, Borgland SL. A role for hypocretin/orexin in motivation. *Behav Brain Res.* 2011; **217**(2): 446-453.
81. Korotkova TM, Sergeeva OA, Eriksson KS, Haas HL, Brown RE. Excitation of ventral tegmental area dopaminergic and nondopaminergic neurons by orexins/hypocretins. *J Neurosci.* 2003; **23**(1): 7-11.
82. Borgland SL, Taha SA, Sarti F, Fields HL, Bonci A. Orexin A in the VTA is critical for the induction of synaptic plasticity and behavioral sensitization to cocaine. *Neuron.* 2006; **49**(4): 589-601.
83. Vittoz NM, Berridge CW. Hypocretin/orexin selectively increases dopamine efflux within the prefrontal cortex: involvement of the ventral tegmental area. *Neuropsychopharmacology.* 2006; **31**(2): 384-395.
84. España RA, Oleson EB, Locke JL, Brookshire BR, Roberts DCS, Jones SR. The hypocretin-orexin system regulates cocaine self-administration via actions on the mesolimbic dopamine system. *Eur J Neurosci.* 2010; **31**(2): 336-348.
85. Borgland SL, Chang S-J, Bowers MS, *et al.* Orexin A/hypocretin-1 selectively promotes motivation for positive reinforcers. *J Neurosci.* 2009; **29**(36): 11215-11225.
86. Teske JA, Levine AS, Kuskowski M, Levine JA, Kotz CM. Elevated hypothalamic orexin signaling, sensitivity to orexin A, and spontaneous physical activity in obesity-resistant rats. *Am J Physiol Regul Integr Comp Physiol.* 2006; **291**(4): R889-R899.
87. Funato H, Tsai AL, Willie JT, *et al.* Enhanced orexin receptor-2 signaling prevents diet-induced obesity and improves leptin sensitivity. *Cell Metab.* 2009; **9**(1): 64-76.
88. Nobunaga M, Obukuro K, Kurauchi Y, *et al.* High fat diet induces specific pathological changes in hypothalamic orexin neurons in mice. *Neurochem Int.* 2014; **78**: 61-66.
89. Zhu Y, Yamanaka A, Kunii K, Tsujino N, Goto K, Sakurai T. Orexin-mediated feeding behavior involves both leptin-sensitive and -insensitive pathways. *Physiol Behav.* 2002; **77**(2-3): 251-257.
90. Horvath TL, Gao X-B. Input organization and plasticity of hypocretin neurons: possible clues to obesity's association with insomnia. *Cell Metab.* 2005; **1**(4): 279-286.
91. Cristino L, Busetto G, Imperatore R, *et al.* Obesity-driven synaptic remodeling affects endocannabinoid control of orexinergic neurons. *Proc Natl Acad Sci U S A.* 2013; **110**(24): E2229-E2238.
92. Teegarden SL, Nestler EJ, Bale TL. Delta FosB-mediated alterations in dopamine signaling are normalized by a palatable high-fat diet. *Biol Psychiatry.* 2008; **64**(11): 941-950.
93. Valdivia S, Patrone A, Reynaldo M, Perello M. Acute high fat diet consumption activates the mesolimbic circuit and requires orexin signaling in a mouse model. *PLoS One.* 2014; **9**(1): e87478.
94. Ahima RS, Flier JS. Leptin. *Annu Rev Physiol.* 2000; **62**: 413-437.

95. El-Haschimi K, Pierroz DD, Hileman SM, Bjørbaek C, Flier JS. Two defects contribute to hypothalamic leptin resistance in mice with diet-induced obesity. *J Clin Invest*. 2000; **105**(12): 1827–1832.
96. Trinko R, Gan G, Gao X-B, Sears RM, Guarnieri DJ, DiLeone RJ. Erk1/2 mediates leptin receptor signaling in the ventral tegmental area. *PLoS One*. 2011; **6**(11): e27180.
97. Hommel JD, Trinko R, Sears RM, et al. Leptin receptor signaling in midbrain dopamine neurons regulates feeding. *Neuron*. 2006; **51**(6): 801–810.
98. Thompson JL, Borgland SL. Presynaptic leptin action suppresses excitatory synaptic transmission onto ventral tegmental area dopamine neurons. *Biol Psychiatry*. 2013; **73**(9): 860–868.
99. Krügel U, Schraft T, Kittner H, Kiess W, Illes P. Basal and feeding-evoked dopamine release in the rat nucleus accumbens is depressed by leptin. *Eur J Pharmacol*. 2003; **482**(1–3): 185–187.
100. van der Plasse G, van Zessen R, Luijendijk MCM, et al. Modulation of cue-induced firing of ventral tegmental area dopamine neurons by leptin and ghrelin. *Int J Obes (Lond)*. In press. DOI: 10.1038/ijo.2015.131.
101. Fulton S, Pissios P, Manchon RP, et al. Leptin regulation of the mesoaccumbens dopamine pathway. *Neuron*. 2006; **51**(6): 811–822.
102. Roseberry AG, Painter T, Mark GP, Williams JT. Decreased vesicular somatodendritic dopamine stores in leptin-deficient mice. *J Neurosci*. 2007; **27**(26): 7021–7027.
103. Matheny M, Shapiro A, Tümer N, Scarpace PJ. Region-specific diet-induced and leptin-induced cellular leptin resistance includes the ventral tegmental area in rats. *Neuropharmacology*. 2011; **60**(2–3): 480–487.
104. Bruijnzeel AW, Qi X, Corrie LW. Anorexic effects of intra-VTA leptin are similar in low-fat and high-fat-fed rats but attenuated in a subgroup of high-fat-fed obese rats. *Pharmacol Biochem Behav*. 2013; **103**(3): 573–581.
105. van den Heuvel JK, Eggels L, Fliers E, Kalsbeek A, Adan RAH, la Fleur SE. Differential modulation of arcuate nucleus and mesolimbic gene expression levels by central leptin in rats on short-term high-fat high-sugar diet. *PLoS One*. 2014; **9**(1): e87729 doi:10.1371/journal.pone.0087729.
106. Powley TL. The ventromedial hypothalamic syndrome, satiety, and a cephalic phase hypothesis. *Psychol Rev*. 1977; **84**(1): 89–126.
107. McGowan MK, Andrews KM, Grossman SP. Chronic intrahypothalamic infusions of insulin or insulin antibodies alter body weight and food intake in the rat. *Physiol Behav*. 1992; **51**(4): 753–766.
108. Figlewicz DP, Evans SB, Murphy J, Hoen M, Baskin DG. Expression of receptors for insulin and leptin in the ventral tegmental area/substantia nigra (VTA/SN) of the rat. *Brain Res*. 2003; **964**(1): 107–115.
109. Pardini AW, Nguyen HT, Figlewicz DP, et al. Distribution of insulin receptor substrate-2 in brain areas involved in energy homeostasis. *Brain Res*. 2006; **1112**(1): 169–178.
110. Liu S, Labouëbe G, Karunakaran S, Clee SM, Borgland SL. Effect of insulin on excitatory synaptic transmission onto dopamine neurons of the ventral tegmental area in a mouse model of hyperinsulinemia. *Nutr Diabetes*. 2013; **3**: e97.
111. Labouëbe G, Liu S, Dias C, et al. Insulin induces long-term depression of ventral tegmental area dopamine neurons via endocannabinoids. *Nat Neurosci*. 2013; **16**(3): 300–308.
112. Bruijnzeel AW, Corrie LW, Rogers JA, Yamada H. Effects of insulin and leptin in the ventral tegmental area and arcuate hypothalamic nucleus on food intake and brain reward function in female rats. *Behav Brain Res*. 2011; **219**(2): 254–264.
113. Könnner AC, Hess S, Tovar S, et al. Role for insulin signaling in catecholaminergic neurons in control of energy homeostasis. *Cell Metab*. 2011; **13**(6): 720–728.
114. Portella AK, Silveira PP, Laureano DP, et al. Litter size reduction alters insulin signaling in the ventral tegmental area and influences dopamine-related behaviors in adult rats. *Behav Brain Res*. 2015; **278**: 66–73.
115. Steculorum SM, Solas M, Brüning JC. The paradox of neuronal insulin action and resistance in the development of aging-associated diseases. *Alzheimers Dement*. 2014; **10**(1 Suppl): S3–S11.
116. Rinaman L. Ascending projections from the caudal visceral nucleus of the solitary tract to brain regions involved in food intake and energy expenditure. *Brain Res*. 2010; **1350**: 18–34.
117. Merchenthaler I, Lane M, Shughrue P. Distribution of pre-pro-glucagon and glucagon-like peptide-1 receptor messenger RNAs in the rat central nervous system. *J Comp Neurol*. 1999; **403**(2): 261–280.
118. Larsen PJ, Tang-Christensen M, Holst JJ, Orskov C. Distribution of glucagon-like peptide-1 and other preproglucagon-derived peptides in the rat hypothalamus and brainstem. *Neuroscience*. 1997; **77**(1): 257–270.
119. Alhadeff AL, Rupprecht LE, Hayes MR. GLP-1 neurons in the nucleus of the solitary tract project directly to the ventral tegmental area and nucleus accumbens to control for food intake. *Endocrinology*. 2012; **153**(2): 647–658.
120. Campos RV, Lee YC, Drucker DJ. Divergent tissue-specific and developmental expression of receptors for glucagon and glucagon-like peptide-1 in the mouse. *Endocrinology*. 1994; **134**(5): 2156–2164.
121. Mietlicki-Baase EG, Ortinski PI, Rupprecht LE, et al. The food intake-suppressive effects of glucagon-like peptide-1 receptor signaling in the ventral tegmental area are mediated by AMPA/kainate receptors. *Am J Physiol Endocrinol Metab*. 2013; **305**(11): E1367–E1374.
122. Wang X-F, Liu J-J, Xia J, Liu J, Mirabella V, Pang ZP. Endogenous glucagon-like peptide-1 suppresses high-fat food intake by reducing synaptic drive onto mesolimbic dopamine neurons. *Cell Rep*. 2015; **12**(5): 726–733.
123. Heppner KM, Perez-Tilve D. GLP-1 based therapeutics: simultaneously combating T2DM and obesity. *Front Neurosci*. 2015; **9**: 92.
124. Chen H, Simar D, Morris MJ. Maternal obesity impairs brain glucose metabolism and neural response to hyperglycemia in male rat offspring. *J Neurochem*. 2014; **129**(2): 297–303.
125. Chan YL, Saad S, Simar D, et al. Short term exendin-4 treatment reduces markers of metabolic disorders in female offspring of obese rat dams. *Int J Dev Neurosci*. 2015; **46**: 67–75.
126. Cao X, Xu P, Oyola MG, et al. Estrogens stimulate serotonin neurons to inhibit binge-like eating in mice. *J Clin Invest*. 2014; **124**(10): 4351–4362.
127. Robert SA, Rohana AG, Shah SA, Chinna K, Wan Mohamud WN, Kamaruddin NA. Improvement in binge eating in non-diabetic obese individuals after 3 months of treatment with liraglutide—a pilot study. *Obes Res Clin Pract*. 2015; **9**(3): 301–304.