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1	Original Manuscript
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3	FEMORAL STRAIN DURING WALKING PREDICTED WITH MUSCLE
4	FORCES FROM STATIC AND DYNAMIC OPTIMIZATION
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36 ABSTRACT

37 Mechanical strain plays an important role in skeletal health, and the ability to accurately 38 and noninvasively quantify bone strain *in vivo* may be used to develop preventive 39 measures that improve bone quality and decrease fracture risk. A non-invasive estimation 40 of bone strain requires combined musculoskeletal – finite element modeling, for which 41 the applied muscle forces are usually obtained from static optimization (SO) methods. In 42 this study, we compared finite element predicted femoral strains in walking using muscle 43 forces obtained from SO to those obtained from forward dynamics (FD) simulation. The 44 general trends in strain distributions were similar between FD and SO derived conditions 45 and both agreed well with previously reported *in vivo* strain gage measurements. On the 46 other hand, differences in peak maximum (ε_{max}) and minimum (ε_{min}) principal strain magnitudes were as high as 32% between FD ($\epsilon_{max}/\epsilon_{min} = 945/-1271 \ \mu\epsilon$) and SO ($\epsilon_{max}/\epsilon_{min}$ 47 48 $= 752/-859 \mu\epsilon$). These large differences in strain magnitudes were observed during the 49 first half of stance, where SO predicted lower gluteal muscle forces and virtually no co-50 contraction of the hip adductors compared to FD. The importance of these results will 51 likely depend on the purpose/application of the modeling procedure. If the goal is to 52 obtain a generalized strain distribution for adaptive bone remodeling algorithms, then 53 traditional SO is likely sufficient. In cases were strain magnitudes are critical, as is the 54 case with fracture risk assessment, then bone strain estimation may benefit by including 55 muscle activation and contractile dynamics in SO, or by using FD when practical. 56

57 KEYWORDS: biomechanics, bone, finite element model, mechanical loading,

58 musculoskeletal model

59

60 INTRODUCTION

61

Bone is a dynamic tissue that exhibits a strong structure-function relationship with its mechanical loading environment. Indeed, physically active individuals tend to accrue more bone mass during growth and development, and better maintain this bone mass throughout adulthood, than their more sedentary counterparts (Parfitt, 1994).

Additionally, the loss of ambulation and habitual muscle loading associated with bed rest or paralysis leads to a rapid and profound loss of bone mineral (Edwards et al., 2013a). In the complete absence of mechanical loading bone reverts to its genetic template, normal in shape and size but lacking distinct characteristics in trabecular microarchitecture, the amount of ossification, and thickness and curvature of the cortical diaphysis (Chalmers and Ray, 1962).

72 The process by which bone senses and responds to mechanical loading is known 73 as functional adaptation, and the mechanical signal that drives this adaptive process is 74 bone strain (Lanyon and Skerry, 2001), or some consequence thereof (i.e., strain energy 75 density, fluid flow, microdamage). An accurate estimation of bone strain during activities 76 of daily living such as walking is therefore integral to understanding the relationship 77 between mechanical loading and skeletal health. In the physiological environment, bone 78 strain is the end result of highly complex loading scenarios (i.e., combined axial, bending, 79 shear, and torsional loading) caused by both gravitational and muscular forces. The 80 resulting bone strain can be quantified *in vivo* using strain gages applied directly to the 81 periosteal surface (Burr et al., 1996); however, the application of strain gages is highly 82 invasive and measurements are limited to only a few, superficial locations. Owing to 83 these limitations, researchers have turned to combined musculoskeletal – finite element

modeling techniques for a more non-invasive estimation of bone strain (Anderson and
Madigan, 2013; Speirs et al., 2007; Vahdati et al., 2014; Viceconti et al., 2012; Wagner et
al., 2010).

87 The concurrent solving of musculoskeletal – finite element models is highly 88 computationally intensive. Methods for estimating muscle forces from musculoskeletal 89 models can require hundreds, thousands, or even millions of iterations within numerical 90 optimization routines (Erdemir et al., 2007), and spending hours or even minutes within 91 each iteration solving a finite element model would incur an impractical amount of 92 computational time. As such, it is most common to use a post-processing technique 93 whereby muscle forces derived from a higher-level rigid multibody simulation are used 94 as boundary conditions for a lower-level elastic model to quantify bone strain (Anderson 95 and Madigan, 2013; Speirs et al., 2007; Vahdati et al., 2014; Viceconti et al., 2012; 96 Wagner et al., 2010). Inherent to a post-processing approach is the assumption that the 97 underlying elastic deformation has no influence on the dynamics of the rigid multibody 98 system. For the calculation of bone strain, this assumption is logical given that bone 99 deformation (Burr et al., 1996) is orders of magnitude lower than that of the 100 musculotendonous units (Fukunaga et al., 2001) and would theoretically have a negligible 101 influence on whole-body motion. 102 The redundancy of the musculoskeletal system allows for an infinite number of

103 muscle force combinations capable of producing the observed joint motions during

104 physical activity (Crowninshield and Brand, 1981). This so-called "force-distribution

105 problem" is typically overcome using numerical optimization procedures (Erdemir et al.,

106 2007). For researchers using combined musculoskeletal – finite element modeling

107 techniques, muscle forces are most frequently predicted using inverse dynamics-based 108 static optimization (Anderson and Madigan, 2013; Speirs et al., 2007; Vahdati et al., 109 2014; Wagner et al., 2010). Static optimization is much less computationally intensive 110 than dynamic optimization, which uses forward dynamics simulation to find optimal 111 motions and controls for a given performance objective, such as tracking an experimental 112 dataset and/or minimizing the metabolic energy expended. Although muscle forces from 113 SO and FD have previously been deemed similar for walking (Anderson and Pandy, 114 2001b), SO has been criticized for lacking explicit time-dependent aspects of muscle 115 force production, and for predicting minimal levels of antagonistic muscle co-contraction 116 (Brand et al., 1994; Collins, 1995), which could potentially have a large influence on 117 overall bone deformation and corresponding strain predictions. 118 The purpose of this study was to quantitatively evaluate finite element predicted 119 periosteal strains at the femur during walking using muscle forces estimated from static 120 and dynamic optimization. To this end, a previously described forward dynamics (FD) 121 simulation of walking was performed using a 3D musculoskeletal model (Fig. 1), and 122 intersegmental joint moments from FD were subsequently used in an inverse-dynamics-123 based static optimization (SO) routine. The muscle forces obtained from FD and SO 124 served as post-possessing inputs to a finite element (FE) model of a femur based on 125 clinical computed tomography (CT) data, and the resulting periosteal strains were 126 compared between FD and SO derived conditions.

128 Musculoskeletal modeling

METHODS

127

129	A 3D musculoskeletal model (Fig. 1c) parameterized to represent a young adult
130	female (i.e., 20 to 35 years) with standing height of 1.65 m and body mass of 61.0 kg was
131	used to simulate walking at 1.25 m/s. The model was conceptually similar to other
132	models used to perform FD gait simulations (Allen and Neptune, 2012; Anderson and
133	Pandy, 2001a) and has been previously described in detail (Miller et al., 2015). Briefly,
134	the model consisted of 10 rigid segments (pelvis, trunk, thighs, shanks, feet, toes)
135	connected at nine joints actuated by 78 Hill-based muscle models (Fig. 1b), including 20
136	muscles per leg that crossed the hip and/or physically connected to the femur. Contact
137	between the feet and the ground was modeled by an array of viscoelastic/Coulomb
138	friction elements on the plantar surfaces of the feet and toe segments. Initial muscle
139	parameters were referenced from a cadaver-based lower limb model (Arnold et al., 2010),
140	which were then adjusted so that joint strength characteristics were similar to average
141	dynamometry data for young adult females (Anderson et al., 2007).
142	Forward dynamics simulation. A simulation of one stride of periodic, bilaterally
143	symmetric walking was performed using a dynamic optimization routine described in our
144	previous work (Miller et al., 2012; Miller et al., 2015) and by others (Allen and Neptune,
145	2012; Umberger, 2010). Briefly, the muscle excitations were parameterized as bimodal
146	signals with two magnitude and four timing parameters per muscle (Fig. 1a). The
147	excitation parameters were optimized to track human experimental gait data (Miller et al.,
148	2014). Specific gait variables included in the tracking cost function were average time
149	series for the pelvis (3D), lumbar (3D), hip (3D), knee (1D), and ankle (1D) angles, the
150	ground reaction force (3D), and the knee adduction moment. To discourage solutions
151	that tracked these data with excessive energy expenditure, the metabolic energy per unit

distance traveled was also calculated (Umberger et al., 2003) and added to the cost
function (see Electronic Supplementary Material for details). A parallel simulated
annealing algorithm (SPAN; (Higginson et al., 2005) was used to systematically adjust
muscle excitation parameters so that the cost function was minimized (Fig. 1d). Muscle
excitation timings for larger muscles were constrained to be similar to normative human
electromyogram timing (Sutherland, 2001).

158 Inverse dynamics based static optimization. An inverse dynamics analysis was 159 performed using data obtained from FD simulation to calculate the intersegmental joint 160 forces and moments. The joint moments and muscle moment arms were used as inputs to 161 a SO problem similar to our previous work (Edwards et al., 2010; Miller et al., 2014), 162 which was solved using the interior-point algorithm in the Matlab Optimization Toolbox. 163 At each time step of the simulated gait cycle, the muscle forces were determined such 164 that (i) all joint moments from the inverse dynamics analysis were reproduced (equality 165 constraint) and (ii) the sum of the squared muscle stresses was minimized (Glitsch and 166 Baumann, 1997). This approach, which is conceptually similar to that of Anderson and 167 Pandy (2001b), was chosen to eliminate differences between muscle forces from FD and 168 SO associated with errors in the collection and processing of experimental data, and the 169 estimation of segment anthropometry. All muscles were modeled as ideal force 170 generators with no contractile or elastic properties because previous studies have 171 suggested adjusting solution boundaries by activation dynamics has a negligible influence 172 on muscle force predictions in walking (Anderson and Pandy, 2001b). 173 *Finite element modeling*

174	A FE model of a full femur was obtained from the VAKHUM database
175	(http://www.ulb.ac.be/project/vakhum/). The native geometry and material properties of
176	the model were based on clinical CT data from a female cadaver (age: 99 yrs, height: 155
177	cm, mass: 55 kg). The CT scan had acquisition setting of 120 kVp and 200 mAs, and
178	images were reconstructed with a slice thickness of 2.7 mm and an in-plane pixel
179	resolution of 0.840 mm. The FE model was comprised of 104,945 linear hexahedral
180	elements with 115,835 degrees of freedom, corresponding to a nominal element edge
181	length of 2.0 mm. Increasing element edge length from 2.0 to 3.0 mm changed femoral
182	displacements, principal stresses, and principal strains by less than 3%, indicating
183	adequate convergence at this refinement.
184	The FE model was first scaled longitudinally to the femoral body of the
185	musculoskeletal model, and then scaled radially assuming bone mass scales to body
186	mass, or length diameter ² \propto body mass (McMahon, 1973), as further justified by the
187	observed correlations between whole-body bone mineral content and body mass (Weiler
188	et al., 2000). Elements were assigned to one of 283 linear-elastic material properties
189	based on relationships between Hounsfield units and apparent density after the integral
190	volumetric bone mineral density of the entire femur was increased by 26% to match that
191	of a young adult female (Keaveny et al., 2010). The density-elasticity relationship was
192	based on uniaxial mechanical testing data of femoral neck trabecular bone (Morgan et al.,
193	2003):
104	E = 6950 = 1.49

194 $E = 6850 \rho_{app}^{1.49}$

195 where *E* is the elastic modulus in MPa, and ρ_{app} is the apparent density in g/cm³; all 196 materials were assigned a Poisson's ratio of 0.3. These material property assignments

have previously illustrated excellent agreement ($r^2=0.91$, RMSE<10%) between

198 experimentally measured and FE-predicted principal strains for cadaveric proximal

199 femora loaded in a stance configuration (Schileo et al., 2007).

201 Central File Exchange (http://www.mathworks.com/matlabcentral/fileexchange/24301-

An affine iterative-closest-point registration procedure available from Matlab

202 finite-iterative-closest-point) was used to align the FE and musculoskeletal model femur

203 into a common local coordinate system. Femoral muscle insertion locations from the

204 musculoskeletal model were then mapped to surface nodes of the FE model. Forces for

205 each of the gluteal muscles (i.e., maximus, medius, and minimus) were equally

206 distributed amongst three separate insertion locations, and the force for the adductor

207 magnus muscle was equally distributed amongst four separate insertion locations (Arnold

208 et al., 2010). The FE model was physiologically constrained at the lateral epicondyle,

209 center of the patellar groove, and the femoral head contact point (Speirs et al., 2007). The

210 hip joint contact force and muscle forces obtained from FD and SO at 10% increments of

211 the gait cycle, from 0% to 100% of stance, as well as the instant of the 1^{st} and 2^{nd} peak

resultant hip joint contact force (JCF1 and JCF2) served as boundary conditions for an

213 implicit FE analysis (Fig. 1e), which was solved using Abaqus/Standard v6.13 (Dassault

214 Systèmes Simulia Corp., Providence, RI). All forces were applied as point loads and

215 resulting strain concentrations were removed from further analysis by discarding nodes

and elements in the immediate vicinity of load application (Polgar et al., 2003).

217 Data reduction

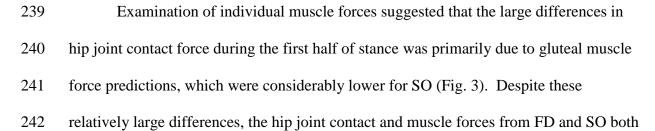
200

The strains occurring along the periosteal surface of the proximal lateral aspect of the femur (35 mm distal to the lateral eminence of the greater trochanter) were compared

220 between FD and SO derived conditions, simply because strains at this location have been 221 directly measured in vivo (Aamodt et al., 1997). To replicate experimental measurement 222 from a strain gage rosette, three-dimensional strains at this location were averaged over a 223 3 x 3 mm region and then transformed into a local coordinate system with a unit normal 224 to the model exterior surface. The longitudinal (ε_{long}), transverse (ε_{trans}), and shear (ε_{shear}) 225 planar strains occurring at this surface were calculated, as well as the maximum (ε_{max}) 226 and minimum (ε_{min}) principal strains, and principal tensile strain (i.e., ε_{max}) angle. For a 227 quantitative comparison of the global femoral strain distribution between FD and SO 228 derived conditions, ε_{max} and ε_{min} occurring along four nodal paths at the anterior, lateral, 229 posterior, and medial periosteal surface of the femoral shaft were quantified at the instant 230 of JCF1 and JCF2.

231 RESULTS

The FD simulation walked at 1.25 m/s with a gross metabolic cost of 3.64 J/m/kg; kinematics and GRF were always within two standard deviations of the experimental means (Fig. 2). Differences in the hip joint contact forces were observed between FD and SO conditions, especially during the first 50% of stance (Fig. 3). While the resultant hip joint contact force at JCF1 was greater in FD than in SO (1824 vs. 1113 N, respectively), the resultant force at JCF2 was quite similar between conditions (1410 vs. 1406 N, respectively).



243 produced femoral bending about an anteriomedial axis with the largest ε_{max} values 244 observed along the lateral surface of the femoral shaft followed by the anterior surface, 245 and the largest ε_{min} values observed along the medial surface followed by the posterior 246 surface (Fig. 4). The unbalanced moments at the patellar groove associated with the 247 physiologic constraints as well as error in the muscle mapping procedure were small and 248 differed only slightly between FD and SO conditions (Fig. 5).

249 The strain predictions at the proximal lateral femur for both FD and SO illustrated 250 a bimodal curve associated with the weight acceptance and push off phases of gait (Fig. 251 6). The largest differences in planar strains were observed at 30% of stance, near the 252 instant of JCF1. At this point in the gait cycle, absolute values of ε_{long} and ε_{trans} , were 253 approximate 37% greater for FD, corresponding to a difference of 278 and 85 μ E, 254 respectively. The absolute difference in $\varepsilon_{\text{shear}}$ at this instant was 7 $\mu\varepsilon$, whereas ε_{max} and 255 ε_{\min} differed by approximate 36%, corresponding to 275 and 84 $\mu\varepsilon$, respectively. 256 Differences in principal strain angle were never greater than 5° . 257 The largest differences in the strain distribution across the length of the femur 258 were observed at JCF1 along the lateral and medial boarders of the femoral shaft (Fig. 7). 259 Peak ε_{max} at JCF1 differed by approximately 250 µ ε along the lateral surface and peak 260 ε_{\min} by 412 µ ε along the medial surface; differences in peak ε_{\max} and ε_{\min} along the 261 anterior and posterior surfaces were relatively small ($\leq 113 \ \mu\epsilon$). Peak ϵ_{max} at JCF2

262 differed by approximately 100 $\mu\epsilon$ along the lateral surface and peak ϵ_{min} by 119 $\mu\epsilon$ along

263 the medial surface. The difference in peak ε_{max} along the anterior surface was 42 $\mu\varepsilon$, while

264 that for peak ε_{\min} along the posterior surface was 171 µ ε .

265 DISCUSSION

266	Mechanical strain plays an important role in skeletal health, and the ability to
267	accurately and non-invasively quantify bone strain in vivo may be used to develop
268	preventive measures that improve bone quality and decrease fracture risk. Our purpose
269	was to compare FE predicted femoral strains during the stance phase of walking using
270	muscles forces obtained from FD simulation and inverse-dynamics-based SO. Despite
271	having identical joint kinematics and intersegmental reaction forces and moments,
272	differences as high as 32% were observed in peak femoral principal strains between FD
273	and SO derived conditions (-1271 $\mu\epsilon$ vs859 $\mu\epsilon$, respectively). The importance of these
274	differences will likely depend on the purpose/application of the combined
275	musculoskeletal – finite element modeling procedure.
276	The muscle force predictions generated by both FD and SO produced femoral
277	bending about an anteriomedial axis with the highest principal tensile strains along the
278	lateral and anterior surface of the femur, and the highest principal compressive strains
279	along the medial and posterior surface of the femur. These global strain distributions are
280	quite similar to those previously reported for FE models loaded with physiological
281	boundary conditions simulating gait (Anderson and Madigan, 2013; Duda et al., 1998;
282	Polgar et al., 2003; Speirs et al., 2007; Wagner et al., 2010). Both FD and SO muscle
283	forces generated peak principal tensile strains and peak principal compressive strains on
284	the order of 500 to 1000 $\mu\epsilon,$ and -1000 to -1500 $\mu\epsilon,$ respectively. These strain magnitudes
285	agree well with previous literature (Duda et al., 1998; Polgar et al., 2003), but in some
286	circumstances are 2-3 times lower (Anderson and Madigan, 2013; Speirs et al., 2007;
287	Wagner et al., 2010). Although some of the discrepancy in strain magnitudes may be
288	associated with specific details of the musculoskeletal model geometry and procedures,

they could just as easily be explained by differences in the calculated/assumed bone mineral density distribution of the femur or material property assignment for the FE model. Near twofold differences in predicted strain magnitudes have been reported between some of the most commonly utilized density-elasticity relationships for FE models of bone (Schileo et al., 2007). Nevertheless, it is important to note that the strain magnitudes observed herein are directly in line with previous *in vivo* measurements (Aamodt et al., 1997; Burr et al., 1996).

Strain gage measurements during walking have been recorded at the proximal 296 297 lateral femur in a 49-year-old female undergoing surgery for "snapping hip syndrome" 298 (Aamodt et al., 1997). The strain gage recordings demonstrated that the proximal lateral 299 femur was undergoing tension during the stance phase of gait. The axial strain along the longitudinal axes of the femur reached 1,133 $\mu\epsilon$, with an ϵ_{max} to ϵ_{min} ratio of -3.05 (1,198/-300 301 393 $\mu\epsilon$), and an average principal tensile strain angle 12° from the longitudinal axis of the 302 femur. The axial strains along the longitudinal axes of the femur for FD reached 745 $\mu\epsilon$ 303 near JCF1, and displayed an ε_{max} to ε_{min} ratio of -3.25 (747/-230 $\mu\epsilon$); corresponding 304 values for SO near JCF1 were: axial strain = 467 $\mu\epsilon$, and ϵ_{max} to ϵ_{min} ratio=-3.22 (471/-305 146 $\mu\epsilon$). The average principal tensile strain angle from the longitudinal axis of the femur 306 during stance was 6.5° (range: -6.0 to 20.6°) for FD and 5.7° (range: -7.3 to 18.8°) for SO. 307 Use of these strain gage data for a rigorous validation of the two modeling procedures 308 employed herein would be a futile exercise, as there are simply too many differences 309 (e.g., age, femoral geometry) and unknowns (e.g., normalcy of gait mechanics following 310 surgery, walking speed, exact location of strain gage) associated with the experimental 311 data. In fact, changing the simulated strain gage location only a few millimeters anterior

312 or inferior increased strain magnitudes by 100 to 200 $\mu\epsilon$. What is important to note is that 313 the general trends in strain for both methods, such as the tension-compression ratio and 314 orientation of the principal axis, seem to correspond with the experimental data. 315 The largest differences in femoral strains between FD and SO were observed 316 during the first half of stance, where SO predicted much lower gluteal muscle forces and 317 virtually no co-contraction of the hip adductors compared to FD. This reduction in 318 muscle co-contraction of the frontal plane hip agonists and antagonist has recently been 319 suggested as a potential cause of lower hip contact force predictions using static 320 optimization when compared to computed muscle control for FD simulation (Wesseling 321 et al., 2015). In this study, differences in muscle forces produced strains that were some 322 30% lower along the lateral and medial surface using SO. On the other hand, peak strain 323 along the anterior surface at JCF1 was higher using SO by nearly 113 μ E. Although joint 324 contact forces were similar between conditions at JCF2, differences in strain distributions 325 up to 171 µɛ were still observed, demonstrating that the relationship between applied load 326 and resulting bone strain is quite complex. A general limitation of this study is that we 327 cannot affirm which method of muscle force estimation is more accurate. Confirmation 328 of accuracy would require *in vivo* measurements of muscle forces thereby negating a

329 musculoskeletal modeling exercise all together. However, practical non-invasive

330 measurements of *in vivo* muscle forces is unlikely to be realized in the near future, and

there remains a need to grow the knowledge base of "best practices" for modeling these

- aspects of human movement (Hicks et al., 2015). Although FD does not necessarily
- 333 predict more realistic muscle forces than SO, it does allow for the prediction of forces
- that are associated with physiologically-motivated objectives (e.g. the propensity to

335 minimize metabolic cost; Srinivasan, 2009) that cannot be included explicitly in SO, and 336 can assess how modeling methods and assumptions may affect the outcomes of 337 simulation studies (Anderson and Pandy, 2001b; Morrow et al., 2014). 338 This study has several limitations that should be borne in mind for the general 339 interpretation and future investigation of combined musculoskeletal – finite element 340 modeling for the non-invasive assessment of bone strain. Although the FE model was 341 modified to have similar size and density to that of a young adult female, the gross 342 morphology and mineral distribution of the model was based on a 99 year-old cadaver. In 343 aging, there is a progressive thinning of the cortical shell (Thompson, 1980) and a 344 reduction in femoral neck shaft angle (Rickels et al., 2011). In fact, the femoral neck shaft 345 angle was approximately 5° lower for the FE model compared to the musculoskeletal 346 model. While these differences may have influenced the absolute values of strain, we 347 have no reason to think that the relative differences between conditions, and thus the 348 interpretation of our findings, would change. This study design allowed for a direct 349 comparison of FD and SO in the absence of experimental error (Anderson and Pandy, 350 2001b). The FD simulation represented only one ensemble average stride of walking, and 351 we can say nothing of the variability of bone strain between strides or different levels of 352 co-contraction. Future work may examine the importance of this variability through 353 stochastic representation of neuromuscular control (Martelli et al., 2015; Viceconti et al., 354 2012).

The FE method is perhaps the most accurate of all current biomechanical modeling approaches because their outputs can be directly compared to *in vitro* experiments (Edwards et al., 2013b; Schileo et al., 2007). Although notably difficult,

358 more work is needed to validate muscle outputs from musculoskeletal models,

359 recognizing that until such time, bone strain predictions based on combined

360 musculoskeletal – finite element modeling may lead to erroneous conclusions regarding

bone factor of safety and remodeling stimuli. If the goal of the modeling procedure is to

362 obtain a generalized strain distribution for adaptive bone remodeling algorithms (Vahdati

363 et al., 2014) both static and dynamic methods should produce analogous results, provided

that stimulus thresholds are adjusted accordingly. In circumstances were strain

365 magnitudes are critical, as is the case with fracture risk assessment (Viceconti et al.,

366 2012), it is possible that these two methods may lead to conflicting conclusions; fatigue-

367 life predictions could potentially differ by one to two orders of magnitude (Carter and

368 Caler, 1985). Performing FD will not always be practical due to the level of modeling and

369 computational effort required, but in these situations, SO routines may benefit by

370 including aspects of muscle force estimation from FD, as is done with methods like

371 computed muscle control (Thelen and Anderson, 2006).

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375 CONFLICT OF INTEREST STATEMENT

376 None of the authors have any conflicts of interest to disclose.

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FIGURE CAPTIONS

Figure 1. Diagram of the combined musculoskeletal – finite element modeling approach for FD simulation. (a) Muscle excitations are defined by six parameters per muscle: two values each for M_i , T_i^{on} , and T_i^{off} . (b) Each of the 78 muscles receives an excitation defined by its six muscle-specific parameters, and develops force in response. (c) Joint moments resulting from muscle forces are applied to the skeleton to cause motion. (d) Excitations are adjusted through optimization to minimize a cost function J. (e) Following optimization, the hip joint contact force and forces for muscles attaching to the femur are used as boundary conditions for a finite element model.

Figure 2. Lumbar, pelvis, and lower limb joint angles and ground reaction forces (GRF) for the FD simulation of walking. Shaded areas are ±2 standard deviations around the mean for 14 human subjects walking in a "normal and comfortable" fashion (Miller et al., 2014). The stride begins and ends at heel-strike. Vertical dashed lines indicate toe-off. The GRF are scaled by bodyweight (BW).

Figure 3. The resultant hip joint contact force and select muscle forces acting at the femur during the stance phase of gait (GMAX=gluteus maximus, GMED=gluteus medius, GMIN=gluteus minimus, ADDMAG=adductor magnus, RECFEM=rectus femoris, VAS = vasti muscles, HAM=hamstring muscles, GAS = gastrocnemius). The hip joint contact force was calculated as the vector sum of the resultant joint reaction force and the forces from muscles spanning the hip.

Figure 4. The minimum principal strain (ε_{min}) distribution on the medial surface (left) and maximum principal strain (ε_{max}) distribution on the lateral surface (right) at JCF1.

Figure 5. The unbalanced moments, or reaction moments, at the distal patellar groove associated with the physiological constraints as well as error in the muscle mapping procedure.

Figure 6. The longitudinal (ε_{long}), transverse (ε_{trans}), and shear (ε_{shear}) planar strains, as well as the maximum (ε_{max}) and minimum (ε_{min}) principal strains, and principal tensile strain (i.e., ε_{max}) angle occurring at the proximal lateral femur (35 mm distal to the lateral eminence of the greater trochanter) as a function of stance.

Figure 7. The maximum principal strains (ε_{max}) along nodal paths of the lateral and anterior surfaces, and minimum principal strains (ε_{min}) along nodal paths of the medial and posterior surfaces at JCF1 (top) and JCF2 (bottom).

FIGURES

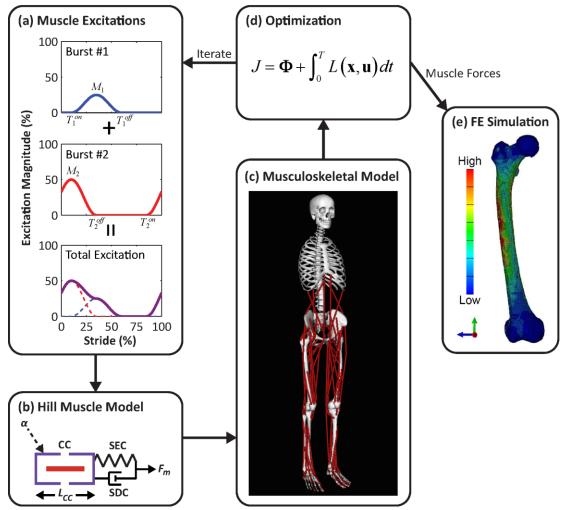


Figure 1.

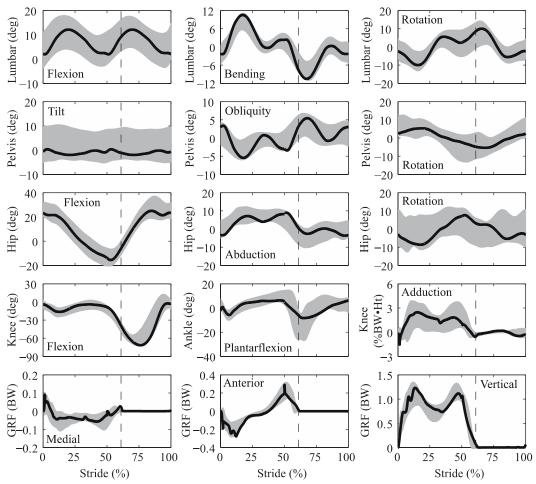


Figure 2.

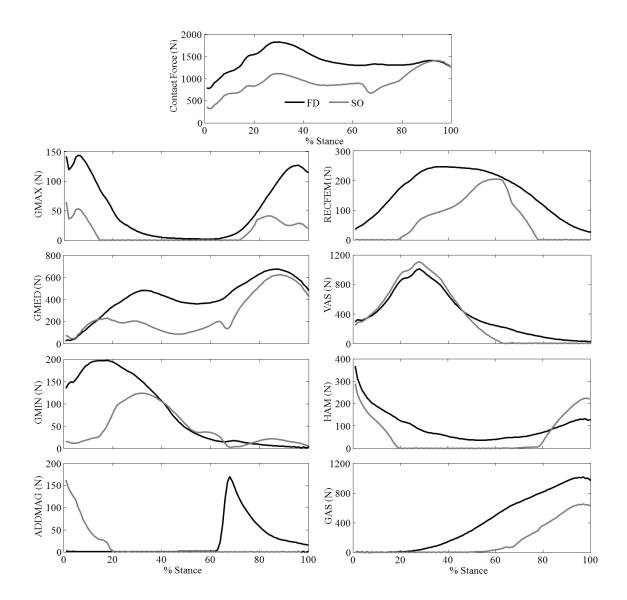


Figure 3.

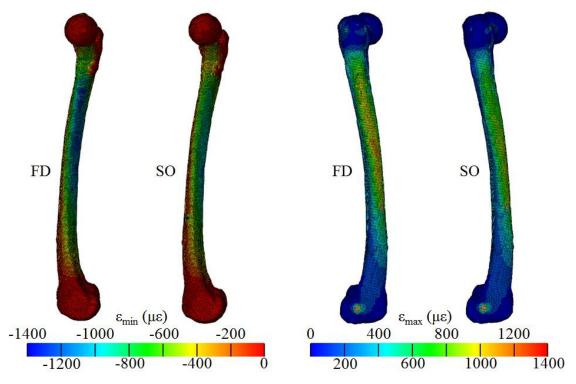


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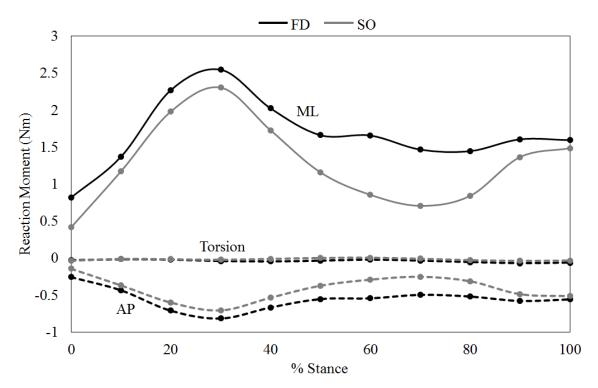


Figure 5.

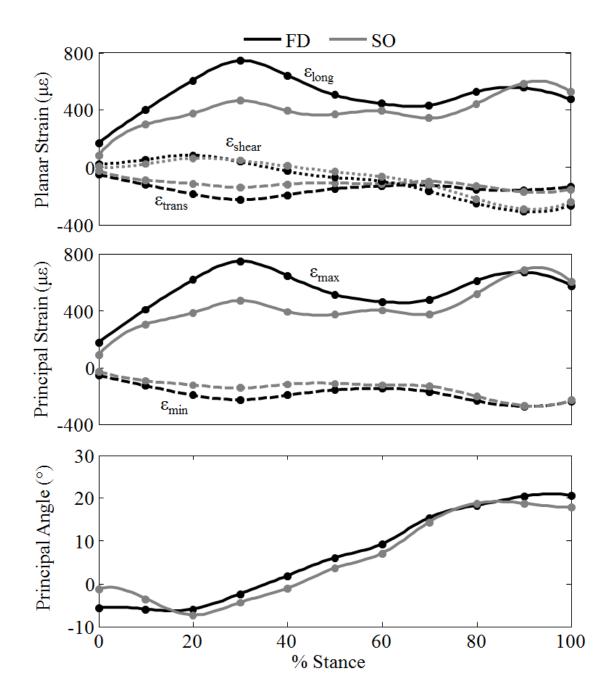
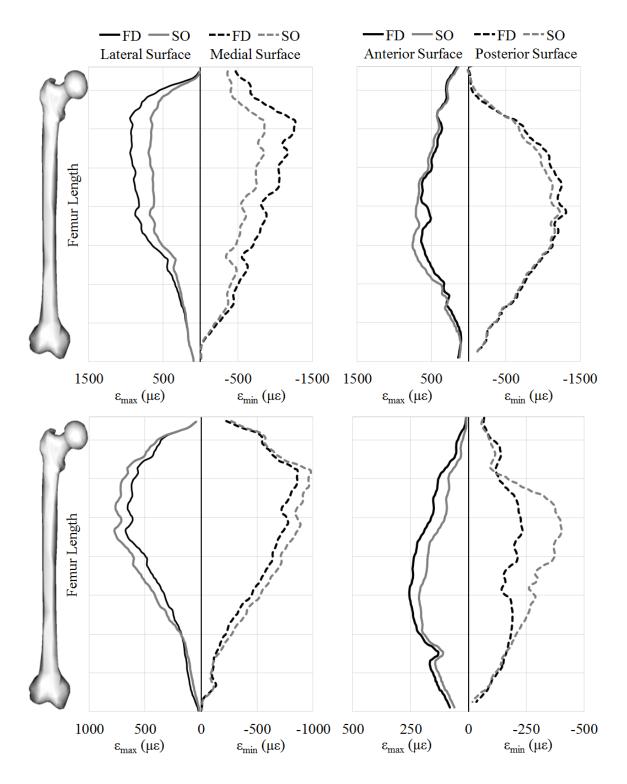


Figure 6.





Electronic Supplementary Material

Femoral strain during walking predicted with muscle forces from static and dynamic optimization

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Cost Function for Forward Dynamics Simulation

The cost function for the forward dynamics simulation was (Miller et al., 2015):

 $J = J_{track} + w_1 J_{metcost} + w_2 J_{exc}$

where J_{track} is the mean squared deviation from the target data, $J_{metcost}$ is the square of the gross metabolic cost of transport (Tucker, 1975), J_{exc} is deviations in the on/off timing of the model's muscle excitations relative to normal human electromyograms (EMG), and w_1 and w_2 are weighting coefficients. The specific form of the tracking function was:

$$J_{track} = \frac{1}{15T} \sum_{i=1}^{15} \left(\int_0^T \left(\frac{x_i^{mod}(t) - x_i^{tar}(t)}{w_i^{tar}(t)} \right)^2 dt \right)$$

where $x_i^{mod}(t)$ is the value of variable *i* at time *t* from the model, $x_i^{tar}(t)$ is the value of the analogous tracking target variable, $w_i^{tar}(t)$ is a weighting factor, and *T* is the step duration. We used means and between-subjects standard deviations for "normal" human walking data from

Miller et al. (2014) to define $x_i^{tar}(t)$ and $w_i^{tar}(t)$, respectively. The 15 tracking targets included the 3D pelvis angles, the 3D lumbar joint angles, the 3D hip angles, the knee flexion angle, the ankle plantarflexion angle, the 3D ground reaction force, and the knee adduction moment. The specific form of the metabolic cost function was:

$$J_{metcost} = \left(\frac{\int_0^T (\dot{E}_{rest} + \sum_{m=1}^{78} \dot{E}_m) dt}{v_{avg} T M}\right)^2$$

where \dot{E}_{rest} is the resting metabolic rate, chosen to be 1.0 W/kg body mass (Waters & Mulroy, 1999), \dot{E}_m is the gross metabolic rate of muscle *m*, calculated using the Umberger et al. (2003) model of human muscle energy expenditure for Hill-based muscle models, v_{avg} is the average horizontal speed of the model's center of mass during $t \in [0,T]$, and *M* is the total body mass. The value of the weighting coefficient $w_1 = 0.2$ was chosen so that a realistic metabolic cost for normal human walking (~ 3.5 J/m/kg at ~ 1.3 m/s; Srinivasan, 2009) had the same weight in the cost function as a reasonably realistic average tracking error for these types of simulations (under ~ two standard deviations on average, e.g. Allen & Neptune, 2012; Miller, 2014).

The specific form of the muscle excitations function was:

$$J_{exc} = \frac{1}{2.78} \sum_{m=1}^{78} \left[(T_i^{on} - \tau_i^{on})^2 + \left(T_i^{off} - \tau_i^{off} \right)^2 \right]$$

where T_i^{on} is the normalized time within the gait cycle (0-100%) when the excitation for muscle *i* turns on, T_i^{off} is the time when this muscle's excitation turns off, and τ_i^{on} and τ_i^{off} are mean on/off off times from indwelling EMG data during healthy adult gait (Sutherland, 2001). The weighting coefficient $w_2 = 50$ increased the cost function score by 0.125 (a very light penalty), 0.5, and 2.0 (a very heavy penalty) for average deviations of 5, 10, and 20% from the normative timing data.

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