



## 21 **Introduction**

22 Hyperkalemic periodic paralysis (HYPP) is a well documented hereditary disease most  
23 commonly associated with Quarter horses descendant from a particular stallion (Aleman  
24 2008). Episodes of HYPP during the peri-operative period have been regularly reported  
25 since the disease was first identified by Cox in 1985 (Cox 1985; Robertson et al. 1992;  
26 Traub-Dargatz et al. 1992; Baetge 2007; Bailey et al. 1996; Cornick et al. 1994; Moody  
27 1995). Of these cases, a limited number have resulted in the successful management of  
28 clinical signs (Cornick et al. 1994; Moody 1995). One previous report has documented  
29 an episode of HYPP during general anaesthesia, following a previous uneventful  
30 anesthetic, though the diagnosis of HYPP was not confirmed (Cornick et al. 1994).  
31 We report the onset of signs leading to the presumed diagnosis and successful  
32 treatment of HYPP in a horse during general anaesthesia. A previous anesthetic, one  
33 week earlier, did not result in an episode of HYPP. The presumptive diagnosis was later  
34 confirmed by DNA analysis.

## 35 **Case Report**

36 A three year old, 400 kg, castrated male Quarter Horse was presented to the Western  
37 College of Veterinary Medicine equine service with a history of epistaxis. Following  
38 endoscopy, a diagnosis of right guttural pouch mycosis involving the internal carotid  
39 artery was made and balloon-tipped catheter occlusion of the artery was planned. Pre-  
40 operative haematology was unremarkable (haematocrit, 27%; total protein, 7.0 g dL<sup>-1</sup>)  
41 and the last observed haemorrhage occurred one week previously. The horse was  
42 bright, alert and responsive and rectal temperature (38.1°C), heart rate (HR; 48 beats  
43 min<sup>-1</sup>) and respiration rate (*f*R; 16 breaths min<sup>-1</sup>) were within normal limits. A cross

44 match was performed with a sample from a donor mare.

45 The horse was fasted overnight and water was available ad libitum. On the morning of

46 surgery the right jugular vein was cannulated (BD Angiocath, 14 GA 5.25 in, Becton

47 Dickinson Infusion Therapy Systems Inc. Sandy, Utah, U.S.A.) and antibiotics (Penicillin

48 G Sodium, 22000 IU kg<sup>-1</sup>, Novopharm, Toronto, Ontario, Canada) and non-steroidal

49 anti-inflammatory drugs (Phenylbutazone, 4.4 mg kg<sup>-1</sup>, 200 mg mL<sup>-1</sup>, Vetoquinol,

50 Lavaltrie, Quebec, Canada) administered intravenously (IV) 30 minutes prior to

51 induction. Premedication consisted of xylazine hydrochloride (1 mg kg<sup>-1</sup>, IV; Rompun,

52 100mg mL<sup>-1</sup>, Bayer (Healthcare), Toronto, Ontario, Canada) and butorphanol tartrate

53 (0.05 mg kg<sup>-1</sup>, IV; Torbugesic, 10mg mL<sup>-1</sup>, Wyeth, Guelph, Ontario, Canada). Following

54 sedation, general anaesthesia was induced with diazepam (0.1 mg kg<sup>-1</sup>, IV; Diazepam,

55 5mg mL<sup>-1</sup>, Sandoz, Burlington, Ontario, Canada) immediately followed by ketamine

56 hydrochloride (2 mg kg<sup>-1</sup>, IV; Ketalean, 100mg mL<sup>-1</sup>, Bimeda-MTC, Animal Health Inc.,

57 Cambridge, Ontario, Canada). Induction and orotracheal intubation (26 mm, ID) was

58 achieved without difficulty. After transfer to the operating theatre and positioning in left

59 lateral recumbency, the horse was connected to a large animal anesthetic machine and

60 anaesthesia maintained with isoflurane (Isoflo, Abbott, Saint Laurent, Quebec, Canada)

61 carried in oxygen. Monitoring equipment was connected and consisted of sidestream

62 capnography, electrocardiography (base-apex configuration; ECG), direct systemic

63 arterial blood pressure (BD Insyte, 20 GA 1.16 in, Becton Dickinson Medical(s) Pte Ltd.,

64 Singapore; placed in the right metatarsal artery), pulse oximetry and rectal temperature.

65 The capnograph was autocalibrated and the pressure transducer equilibrated with

66 atmospheric pressure prior to use. Physiologic parameters were displayed and recorded

every five minutes (Cardiocap/5, Datex-Ohmeda, Louisville, CO, U.S.A.). The anaesthetic period was uneventful: mean direct systemic arterial blood pressure (mABP) ranged from 60 – 90 mmHg (8 - 12 kPa), partial pressure of end-tidal carbon dioxide ( $P_{ET}CO_2$ ) ranged from 36 – 49 mmHg (4.8 - 6.5 kPa), and heart rate ranged from 35-55 beats  $min^{-1}$ . Dobutamine (Dobutamine, 12.5 mg  $mL^{-1}$ , Sandoz (Canada Inc.), Burlington, Ontario, Canada) was administered intravenously as required to support cardiovascular function and ventilation was controlled with intermittent positive pressure ventilation (IPPV; tidal volume [ $V_T$ ], 4 L;  $f_R$  7 – 10 breaths  $min^{-1}$ ). End-tidal isoflurane concentration was 1.3 % for the majority of the procedure. Lactated Ringer's Solution (LRS; Lactated Ringer's, Hospira (Healthcare Corporation), Montreal, Quebec, Canada) was administered IV at a rate of 10 mL  $kg^{-1} hr^{-1}$  through a second IV cannula in the right medial saphenous vein. One arterial blood sample was taken using an anaerobic technique thirty minutes after induction of anaesthesia and analysed immediately (Table). Surgery proceeded without complication and estimated blood loss was less than 50 mL. The duration of the anesthetic period was 2.5 hours. Recovery to standing was smooth and rapid (20 minutes). Xylazine (0.25 mg  $kg^{-1}$ , IV) was administered prior to transfer to the recovery box.

Seven days following surgery, the animal experienced an episode of acute epistaxis in its stall. Bedding in the stall prevented an accurate estimation of blood loss though the total volume was estimated as less than 2 L. The animal was agitated and difficult to approach. Acepromazine (0.025 mg  $kg^{-1}$ , IV), xylazine (0.4 mg  $kg^{-1}$ , IV) and butorphanol (0.04 mg  $kg^{-1}$ , IV) were administered prior to calling the anaesthesia service. Due to the presence of large blood clots, a thorough evaluation of the right guttural pouch had not

90 been possible at the time of the initial endoscopic evaluation. The episode of epistaxis  
91 after occlusion of the internal carotid artery was attributed to involvement of the external  
92 carotid or maxillary artery. Therefore the decision was taken to ligate the right external  
93 maxillary artery and occlude the right major palatine artery with a balloon-tipped  
94 catheter. The right jugular vein was cannulated (14 GA 5.25 in) and a blood sample sent  
95 for haematology (haematocrit, 26%; total protein 5.9 g dL<sup>-1</sup>). Anaesthesia was induced  
96 with diazepam (0.1 mg kg<sup>-1</sup>, IV) immediately followed by ketamine (2 mg kg<sup>-1</sup>, IV) and  
97 orotracheal intubation performed without difficulty (26 mm ID). The animal was  
98 positioned in left lateral recumbency, connected to a large animal anesthetic machine  
99 and IPPV initiated ( $V_T$ , 4 L;  $f_R$ , 8 breaths min<sup>-1</sup>; peak inspiratory pressure [PIP], < 20  
100 cmH<sub>2</sub>O). Anaesthesia was maintained with isoflurane from a precision vaporizer in  
101 oxygen, and the end-tidal isoflurane was between 1.2 – 1.3% throughout the anesthetic  
102 period. Instrumentation and monitoring was as for the first anaesthesia. A whole blood  
103 transfusion had been started pre-operatively and was continued following transfer to the  
104 operating theatre (approximately 1mL kg<sup>-1</sup> hr<sup>-1</sup>). Lactated Ringer's solution was  
105 administered through a separate IV cannula (right medial saphenous vein) at 10 mL kg<sup>-1</sup>  
106 min<sup>-1</sup>. During the anesthetic mABP ranged from 55 – 80 mm Hg (7.3 - 10.7 kPa). The  
107 first arterial blood gas analysis (ABG 1; Table) taken approximately 40 minutes after  
108 induction showed a respiratory acidosis and  $V_T$  was increased to 5L. Respiratory rate  
109 was increased to 10 breaths min<sup>-1</sup> following a second ABG (ABG 2; Table) 40 minutes  
110 later. Up to this point mABP varied frequently between 55 – 70 mmHg (7.3 - 9.3 kPa)  
111 and heart rate varied between 50 – 85 beats min<sup>-1</sup>. Dobutamine was infused  
112 continuously between 0.2 – 0.4 µg kg<sup>-1</sup> min<sup>-1</sup>. Systolic (sABP) and diastolic (dABP) direct

113 systemic arterial blood pressures ranged between 80 – 100 mmHg (10.7 - 13.3 kPa)  
114 and 45 – 60 mmHg (6.0 - 8.0 kPa) respectively. Administration of butorphanol (0.04 mg  
115 kg<sup>-1</sup>, IV) was associated with a decrease in HR (from 85 to 65 beats min<sup>-1</sup>), and mABP  
116 increased from 60 to 75 mm Hg (8.0 - 10.0 kPa) following IV ephedrine (0.05 mg kg<sup>-1</sup>;  
117 administered 15 minutes after butorphanol). A third blood sample (ABG 3; Table)  
118 showed a decrease in PaCO<sub>2</sub> and a slight increase in potassium (from 3.6 mmol L<sup>-1</sup>  
119 [ABG1] to 4.2 mmol L<sup>-1</sup>; reference range 3.5 – 5 mmol L<sup>-1</sup>).

120 Approximately 45 minutes after the previous ABG, an increase in P<sub>ET</sub>CO<sub>2</sub> from 52 to 61  
121 mmHg (6.9 - 8.1 kPa) was noted in addition to muscle fasciculations over the animal's  
122 flanks and a sinus tachycardia (Fig. 1a) of 78 beats min<sup>-1</sup> (mABP was 70 mm Hg [9.3  
123 kPa]). Additionally, the ECG showed a narrowed T wave with increased amplitude (Fig.  
124 1a) Ventilatory parameters were unchanged (V<sub>T</sub>, 4L; f<sub>R</sub>, 8; PIP < 20 cm H<sub>2</sub>O), the  
125 vaporizer had an adequate amount of anesthetic agent and the depth of anaesthesia  
126 appeared unchanged from examination of palpebral reflexes and neck muscle tone. The  
127 partial pressure of inspired CO<sub>2</sub> was negligible (less than 2 mm Hg [0.3 kPa]). Rectal  
128 temperature was unchanged, at 35.5 °C. An ABG (ABG 4, Table) taken at this time  
129 revealed a serum potassium concentration of 7.2 mmol L<sup>-1</sup> and PaCO<sub>2</sub> of 78 mm Hg  
130 (10.4 kPa).

131 Hyperkalaemic periodic paralysis was suspected on the basis of clinical signs, ECG  
132 changes, ABG results and breed. Over the next 10 minutes respiratory rate was  
133 increased to 12 breaths min<sup>-1</sup> and calcium gluconate (0.1-0.2 mg kg<sup>-1</sup> min<sup>-1</sup>; Calcium  
134 gluconate, 23%, Calgary, Alberta, Canada) and dextrose 5% in saline (5 mL kg<sup>-1</sup> hr<sup>-1</sup>;  
135 Dextrose, 50%, Bimeda-MTC, Animal Health Inc., Cambridge, Ontario, Canada) were

136 administered through two separate IV cannulae. Fluid therapy was changed from LRS  
137 to NaCl 0.9% and the whole blood transfusion was stopped. A second dose of  
138 ephedrine was given ( $0.05 \text{ mg kg}^{-1}$ , IV) as mABP had decreased to 55 mm Hg (7.3 kPa).  
139 The mABP increased to 85 mm Hg (11.3 kPa) shortly afterwards and remained above  
140 this level for the remainder of the anesthetic. Approximately 10 minutes after IV calcium  
141 gluconate administration had been started, the ECG showed T waves with normal  
142 morphology (Fig. 1b; complex six is a premature complex of atrial or junctional origin).  
143 Subsequent ABGs (ABG 6 and 7, Table) showed a gradual decrease in  $\text{PaCO}_2$ ,  $\text{HCO}_3^{-1}$ ,  
144 and potassium, although these parameters remained supranormal. The surgical  
145 procedure was successful and terminated one hour after signs associated with HYPP  
146 were first observed. The animal was transferred to the recovery box and romifidine ( $0.01$   
147  $\text{mg kg}^{-1}$ ; Sedivet,  $10 \text{ mg mL}^{-1}$ , Boehringer Ingelheim, Burlington, Ontario, Canada)  
148 administered IV. Anesthetic duration was 4 hours. The total volumes of fluids  
149 administered were: LRS, 15L; NaCl 0.9%, 3L; dextrose 5%, 2L; whole blood, 1.5L.  
150 Intravenous dextrose and normal saline were continued during recovery. Regular (NPH)  
151 insulin ( $0.05 \text{ IU kg}^{-1}$ , subcutaneously; HumulinN NPH,  $100 \text{ U mL}^{-1}$ , Lilly, Toronto, Ontario,  
152 Canada) was administered in recovery box. Unassisted recovery to standing was  
153 uneventful and completed in 45 minutes. A jugular venous blood sample (VBG, Table)  
154 taken after standing showed a return to normal serum potassium levels. Analysis of the  
155 whole blood intended for transfusion revealed a potassium concentration within  
156 reference range (Table). Several hours later, a jugular venous blood sample showed a  
157 normal potassium concentration ( $3.8 \text{ mmol L}^{-1}$ ) and the horse showed no signs of  
158 muscle fasciculations or weakness. Questioning of the owner confirmed genetic lineage

to a known carrier of HYPP and a blood sample submitted for DNA analysis confirmed a diagnosis of HYPP (heterozygous).

## **Discussion**

Since the initial reports of HYPP, further investigation identified an autosomal dominant mode of inheritance, resulting in abnormal sodium channels in skeletal muscle (Cox 1985; Naylor et al. 1992; Pickar et al. 1991; Spier et al. 1993; Steiss & Naylor 1986).

The defective alpha subunit gene and resultant amino acid substitution (leucine for phenylalanine) form sodium channels that fail to inactivate following depolarisation, causing a reduction (more positive) in the resting membrane potential (RMP) as a result of the increase in sodium permeability (Rudolph et al. 1992a, b). The reduced RMP allows potassium to diffuse out of the cell, potentially leading to hyperkalaemia. As the RMP approaches the threshold potential, potassium efflux increases, eventually leading to clinical signs of HYPP (Aleman 2008; Meyer et al. 1999). The incidence of heterozygous or homozygous carriers within the Quarter Horse population is between 0.4 and 4% (Aleman 2008; Naylor 1994). Factors associated with the development of a clinical episode of HYPP in horses include stress, diets high in potassium, anaesthesia, transportation and concurrent disease (Meyer et al. 1999). In this case, possible reasons why the second anesthetic resulted in clinical symptoms whilst the first did not, include the occurrence of pre-operative stress and agitation associated with sudden haemorrhage, high levels of activity and handling within the stall, and the hypercapnia which developed soon after induction, prior to an elevation in serum potassium. Prior to potassium measurement the whole blood transfusion was considered a source of external potassium as blood products may contain high levels of potassium resulting

from hemolysis (Hohenhaus 2006). From published reports of peri-operative HYPP, common clinical signs include elevated  $P_{ET}CO_2$  and hypercapnia, muscle fasciculations and hyperkalaemia associated with altered T wave morphology (narrowing and increased amplitude) (Baetge 2007; Bailey et al. 1996; Cornick et al. 1994; Moody 1995; Robertson et al. 1992; Traub-Dargatz et al. 1992). Many of these signs overlap with those of the hypermetabolic syndrome malignant hyperthermia (MH; hyperthermia, muscle contractions and rigidity, hyperkalaemia, hypercapnia, tachypnea, tachycardia, systemic hypertension, arrhythmias, sudation), which has been confirmed and suspected in the horse (Aleman et al. 2005, 2009; Klein et al. 1989). Compared with the other reports of confirmed/ suspected HYPP during the perioperative period, Cornick et al. (1994) reported a suspected case which exhibited repeated hyperthermia during several anesthetics, peaking at a body temperature of 40.2°C.

In the case reported here the differentiation between MH and HYPP was based on the stability of rectal temperature, and absence of sudation and muscle rigidity. Similar to our case, previous reports have reported tachycardia during an episode of HYPP (Bailey et al. 1996; Cornick et al. 1994). In conscious horses, hyperkalaemia results in bradycardia and a progressive deterioration of the cardiac rhythm as the RMP approaches and eventually exceeds the threshold potential. The ECG changes mirror serum potassium concentration, progressing from a widening and flattening of P waves, to an increase in P-R interval, S-T segment elevation, increased T wave amplitude, widening of the QRS complex and eventual ventricular fibrillation (Meyer et al. 1999; Spier et al. 1990). This apparent paradox of tachycardia in the face of hyperkalaemia may reflect the opposing effects of hyperkalaemia and hypercapnia on heart rate as



205 these reports also reported the concurrent presence of hypercapnia. The site of muscle  
206 fasciculation varies between reports and the following sites have been described:  
207 thoracic limb alone, all limbs, head, generalized (Baetge 2007; Bailey et al. 1996;  
208 Cornick et al. 1994). As reported by Bailey et al. (1996) and Baetge (2007), we also  
209 found that the hypercapnia was difficult to control despite IPPV. Recorded levels of  
210 serum potassium associated with HYPP in the perioperative period range from 3.5 – 8.2  
211 mmol L<sup>-1</sup> (Baetge 2007; Bailey et al. 1996; Cornick et al. 1994; Moody 1995; Robertson  
212 et al. 1992).

213 Recommended treatment for clinical cases of HYPP aim to address the hyperkalaemia,  
214 associated arrhythmia and respiratory acidosis. This includes administration of  
215 potassium-free fluids, a source of intravenous calcium and glucose, increased minute  
216 ventilation to reduce PaCO<sub>2</sub>, and insulin (Bailey et al. 1996; Meyer et al. 1999). The  
217 administration of sodium bicarbonate is occasionally advocated as alkalinization of the  
218 blood results in an exchange of potassium for hydrogen ions, lowering serum  
219 potassium. However, the potential benefits of sodium bicarbonate are tempered by  
220 numerous risks, including hypotension associated with rapid administration, a decrease  
221 in myocardial contractility resulting from a decrease in ionised calcium concentration,  
222 and iatrogenic metabolic alkalosis. The administered fluid was changed from LRS to  
223 0.9% NaCl in an effort to limit any further contribution to serum potassium though this  
224 approach has been recently questioned in hyperkalemic cats with urinary obstruction  
225 (Drobatz 2008). Intravenous calcium gluconate is useful for limiting the arrhythmic  
226 effects of hyperkalaemia by raising the threshold potential for the generation of action  
227 potentials. As it does not address the hyperkalaemia directly, specific treatments for

228 lowering plasma potassium are necessary, including glucose and insulin. The effect of  
229 glucose is to stimulate insulin release, which drives cellular uptake of glucose and  
230 potassium, resulting in a lowering of serum potassium concentration. Insulin has an  
231 additional effect of activating the Na<sup>+</sup>-K<sup>+</sup> ATPase pump, increasing cellular uptake of  
232 potassium ions. Efforts to reduce PaCO<sub>2</sub> by an increase in minute ventilation were  
233 partially successful. Hypercapnia results in an increase in serum potassium levels,  
234 further contributing to hyperkalaemia. At the end of the surgery, potassium levels  
235 remained elevated (ABG 7, Table 1) and a low dose of insulin was administered upon  
236 transfer to recovery. The horse was left to recover unassisted in an effort to minimize  
237 any further stress provoking a second episode of HYPP. An attempt to limit the risk of  
238 administering insulin, during a period when blood glucose monitoring was not possible,  
239 was made by administering intravenous dextrose (in normal saline) until the horse  
240 started to move.

241 Our case differs from that of Bailey et al. (1996) in the therapy pursued. In addition to  
242 calcium gluconate, dextrose and insulin were administered. The combined effects were  
243 a reduction in serum potassium concentration to within the normal range shortly after  
244 the end of the anesthetic. This may have contributed to the smooth recovery observed  
245 and absence of muscle fasciculations. Additionally, the whole blood transfusion was  
246 stopped as a potential source of potassium (Hohenhaus 2006). With the exception of  
247 insulin administration, all treatment had been initiated in the operating theatre. The slow  
248 response to therapy, in terms of serum potassium levels (ABG 6 and 7), likely reflect the  
249 short duration over which the treatments were administered. Only 3L of 0.9% NaCl were  
250 administered during this time and treatment was directed to ensuring the adequate

administration of calcium and dextrose, which were considered more important in terms of correcting the arrhythmia.

This case was similar to that reported by Cornick et al. (1994) in that an initial anaesthetic period did not result in clinical signs of HYPP or an elevation in serum potassium concentrations. Of the cases reported in the literature, diagnosis was confirmed by DNA analysis in two cases (Bailey et al. 1996; Moody 1995). Our case differs from others in the unique combination of the following factors: signs of HYPP became evident intra-operatively following a previous uneventful general anesthetic using an identical anesthetic protocol. Diagnosis was rapidly achieved and aggressive treatment instituted promptly, resulting in the resolution of clinical signs and an unassisted, uneventful recovery. The diagnosis was later confirmed by DNA analysis. Following a previous (1998) recommendation for testing of foals with known HYPP ancestry (related to the stallion Impressive), the American Quarter Horse Association has made testing a requirement for all foals born after 1<sup>st</sup> January, 2007 with known HYPP ancestry.

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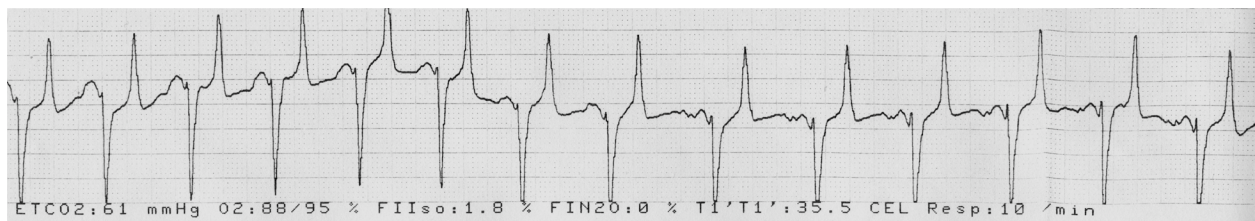
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Table. Blood-gas and electrolyte results from the first and second surgeries.

ABG = arterial blood-gas and electrolytes. VBG = venous blood-gas and electrolytes.

\*start of treatment for suspected HYPP. †sample taken when horse recovered to standing following anaesthesia.

Fig. 1a: Base-apex ECG recorded prior to therapy for suspected HYPP. Sinus tachycardia of 78 beats  $\text{min}^{-1}$  and tall, narrow T waves exhibited. Paper speed, 25 mm  $\text{sec}^{-1}$ . Height 0.5 mV per division.



337 Fig. 1b: Base-apex ECG recorded following initiation of therapy for suspected HYPP.  
338 Return of normal T wave morphology. Heart rate of 59 beats min<sup>-1</sup>. Complex six is a  
339 premature complex of atrial or junctional origin. Paper speed, 25 mm sec<sup>-1</sup>. Height 0.5  
340 mV per division.

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