

1 Effects of cyclic loading on the mechanical properties and failure of human patellar tendon

2

3 Colin R. Firminger^{1,2,3}, W. Brent Edwards^{1,2,3}

4

5 ¹Human Performance Laboratory, Faculty of Kinesiology, University of Calgary, Canada

6 ²Biomedical Engineering Graduate Program, University of Calgary, Canada

7 ³McCaig Institute for Bone and Joint Health, University of Calgary, Canada

8

9 **Corresponding Author:** Colin Firminger

10 **Mailing Address:** KNB 219, Human Performance Laboratory, University of Calgary, 2500

11 University Drive NW, Calgary, AB Canada, T2N 1N4

12 **Phone:** 403-808-2601

13 **Email:** cfirring@ucalgary.ca

14

15 **Word Count:** 3734

16

17 **Keywords:** Fatigue loading; Tendinopathy; Digital image correlation; Tendon mechanics; Surface

18 strain distributions

19 **Abstract**

20 Patellar tendinopathy is a common overuse injury in sports such as volleyball, basketball, and long-
21 distance running. Microdamage accumulation, in response to repetitive loading of the tendon,
22 plays an important role in the pathophysiology of patellar tendinopathy. This damage presents
23 mechanically as a reduction in Young's modulus and an increase in residual strain. In this study,
24 19 human patellar tendon samples underwent cyclic testing in load control until failure, segmented
25 by four ramped tests where digital image correlation (DIC) was used to assess anterior surface
26 strain distributions. Ramped tests were performed prior to cyclic testing and at timepoints
27 corresponding to 10%, 20%, and 30% of cyclic stiffness reduction. Young's modulus significantly
28 decreased and cyclic energy dissipation significantly increased over the course of cyclic testing.
29 The DIC analysis illustrated a heterogeneous strain distribution, with strain concentrations
30 increasing in magnitude and size over the course of cyclic testing. Peak stress and initial peak
31 strain magnitudes significantly correlated with the number of cycles to failure ($r^2=0.65$ and
32 $r^2=0.57$, respectively, $p<0.001$); however, the rates of peak cyclic strain and modulus loss
33 displayed the highest correlations with the number of cycles to failure ($r^2=96\%$ and $r^2=86\%$,
34 respectively, $p<0.001$). The high correlation between the rates of peak cyclic strain and modulus
35 loss suggest that non-invasive methods to continuously monitor tendon strain may provide
36 meaningful predictions of overuse injury in the patellar tendon.

37 **Introduction**

38 Patellar tendinopathy is a common overuse injury observed in athletes participating in sports
39 characterized by repetitive loading and cumulative bouts of activity (Cook et al., 1997; Ferretti et
40 al., 1984; Lian, 2005; Malliaras et al., 2015). Patellar tendinopathy has an insidious onset and a
41 prolonged recovery time, forcing many athletes to retire early from sport (Cook et al., 1997;
42 Kettunen et al., 2002; Malliaras et al., 2015). Tendinopathy is associated with degenerative
43 changes including collagen disorganization, hypercellularity, and hypervascularization (Kannus
44 and Jozsa, 1991; Tallon et al., 2001), as well as reduced Young's modulus (Arya and Kulig, 2010).
45 The pathophysiology of tendinopathy remains a strong topic of debate (Millar et al., 2017), but
46 whether a purely mechanically or biologically mediated phenomenon, tendon damage associated
47 with repetitive loading plays a decided role in this injury (Fung et al., 2010).

48

49 The effects of fatigue loading have been observed at several length scales within the extracellular
50 matrix, including collagen molecule denaturation (Zitnay et al., 2020), fibril sliding (Lee and
51 Elliott, 2019), kinked fibers (Szczesny et al., 2018), and complete matrix disruptions (Fung et al.,
52 2010; Gibbon et al., 1999; Nakama et al., 2005). The degradation of collagen in response to fatigue
53 loading causes material property deterioration in the form of reduced tissue modulus and ultimate
54 tensile strength, along with increased cyclic energy dissipation and peak cyclic strain. (Fung et al.,
55 2010; Schechtman and Bader, 2002; Wren et al., 2003). While the exact relationship between
56 microdamage accumulation and tissue mechanical properties is complex, quantifying the effects
57 of cyclic loading on tendon mechanical properties may be used to predict the fatigue failure of
58 tendon.

59

60 The influence of cyclic loading on the fatigue behaviour of tendon has been examined in several
61 animal models (Fung et al., 2010; Ker et al., 2000; Pike et al., 2000; Thorpe et al., 2015; Wang et
62 al., 1995); however, research examining human tendon is limited (Schechtman and Bader, 2002,
63 1997; Wren et al., 2003). Previous studies of human tendon performed *in vitro* cyclic tests of
64 cadaveric extensor digitorum longus (Schechtman and Bader, 2002, 1997) and Achilles (Wren et
65 al., 2003) tendons, reporting significant exponential relationships between fatigue life (i.e., the
66 number of cycles to failure) and both peak stress and initial peak strain. While these venerable
67 studies highlighted the important relationship between peak stress/strain magnitude and the fatigue
68 life of tendon, specific details of this relationship are likely to differ amongst different anatomical
69 locations, as the fatigue quality of tendon appears to be highly adapted to the stresses experienced
70 during daily activity (Ker et al., 2000; Pike et al., 2000).

71
72 Previous studies utilizing optical methods to track tendon displacement at the tissue level
73 illustrated that tissue strains are location-dependent (Wren et al., 2003). Digital image correlation
74 (DIC) is a state-of-the-art technique that has been used to non-invasively quantify strains across
75 the entire surface of a material by tracking the displacement of a stochastic speckle pattern (Luyckx
76 et al., 2014; Palanca et al., 2016). By integrating this technique into a fatigue test, spatiotemporal
77 changes in strain can be visualized, potentially providing insight into the failure mechanism for
78 tendons under cyclic loading.

79
80 Understanding how cyclic loading alters the material properties of human patellar tendon may be
81 used to identify pertinent metrics that are associated with fatigue failure and create tools to monitor
82 the risk of developing patellar tendinopathy. Therefore, the purpose of this study was to quantify

83 the fatigue behaviour of human patellar tendon using nominal and DIC-based measures. More
84 specifically, we sought to evaluate changes in tissue modulus, cyclic energy dissipation, and DIC-
85 based anterior surface strains during cyclic loading, and to establish correlations between fatigue
86 life with measures of stress, strain, and time-dependent measures such as the rate of peak cyclic
87 strain increase (i.e., creep rate) and Young's modulus degradation rate (i.e., damage rate). We
88 hypothesized cyclic loading would decrease tissue modulus and increase cyclic energy dissipation
89 and surface strains over the course of the fatigue test, and that fatigue life would be well described
90 by non-linear relationships with stress, strain, creep rate, and damage rate.

91

92 **Methods**

93 *Specimen Preparation and Test Setup*

94 Thirty fresh-frozen cadaveric knees (mid-thigh to mid-shank) were obtained from Science Care
95 (AZ, USA). Cadaveric testing was approved by the University of Calgary's Conjoint Health
96 Research Ethics Board (REB18-0970). Prior to dissection, specimens were thawed for 12 hours in
97 a plastic bag submerged in lukewarm water followed by computed tomography (CT) imaging
98 (Revolution HD GSI; GE Healthcare, IL, USA) to examine for any obvious joint pathology. Scans
99 were performed with a peak voltage of 120 kVp, a tube current of 220 mA, with an in-plane
100 resolution = 0.39×0.39 mm and a slice thickness of 0.63 mm. Next, the patellar tendon (attached
101 to the patella and tibia) was dissected stored in a -20°C freezer for a maximum of 6 months.

102

103 Prior to mechanical testing, each specimen was thawed for 45 minutes at room temperature while
104 fully submerged in saline. A handheld reciprocating bone saw (M.SR-SCT; Freedom, CT, USA)
105 was then used to cut the tibial diaphysis 40 mm inferior to the patellar tendon enthesis and to

106 remove the tibial plateau (Figure 1a). The medial, lateral, and posterior aspects of the tibia between
107 the cut and the patellar tendon enthesis were removed to create a rectangular shape that could be
108 held in a vertical clamp without touching the patellar tendon enthesis. The medial and lateral
109 portions of the patellar tendon were removed by cutting parallel to the tendon fascicles with a
110 scalpel (Figure 1a). This was necessary to reduce the force required to produce a given tendon
111 stress, as pilot testing identified that the bony endpoints occasionally fractured at high loading
112 magnitudes.

113
114 The patellar tendons were inverted and placed into two custom made clamps (Figure 1b) with both
115 entheses outside of the clamped region. The lower clamp was rotated to create a patella-patellar
116 tendon angle of 146° ($180^\circ = \text{straight}$), corresponding to the average knee angle at peak tendon
117 strain during a maximal countermovement jump (Firminger et al., 2019; Schmid et al., 2002). A
118 laser level (DW088K; Dewalt , MD, USA) was used to ensure the tendon was vertical, and a matte
119 black acrylic speckle pattern was painted on to the tendon's anterior surface using an airbrush with
120 a 0.3 mm tip (Master Precision G444; Master Airbrush, China). Gauge length was measured with
121 digital calipers as the distance between the patellar and tibial entheses. Cross-sectional area was
122 calculated assuming a rectangular cross-section as this best represented the tendon geometry
123 following the preparation protocol. Cross-sectional area was measured using digital calipers by
124 averaging the results from three width and three thickness measurements taken halfway along the
125 gauge length.

126

127 *Mechanical Testing*

128 Tendon samples were cyclically loaded until failure (i.e., full tendon rupture), segmented by four
129 ramped loading tests to capture images for DIC analysis (Figure 2). An ElectroPuls E10000
130 materials testing machine was used for all mechanical tests (Instron, MS, USA). Samples were
131 kept moist using four saline drips positioned on the tendon, each with a flow rate of $0.2 \text{ mL} \cdot \text{min}^{-1}$
132 ¹. The selected flow rate was selected based on pilot testing, and ensured that the sample was
133 visibly hydrated throughout the testing procedure (i.e., excess saline was dripping from the
134 tendon).

135
136 Cyclic testing was performed in load control using a 2 Hz sinusoidal waveform. The tendons were
137 loaded between 2 MPa and a randomly assigned peak stress, ranging between 11% and 60% of the
138 estimated 60 MPa ultimate tensile strength (UTS) for patellar tendon (Hashemi et al., 2005;
139 Johnson et al., 1994; Noyes et al., 1984). This range of stress values was selected to span estimated
140 patellar tendon loads during walking and jogging (Firminger and Edwards, 2016; Lee and Hidler,
141 2008). Data were captured at 200 Hz for every cycle until the 100th cycle, then every 10th cycle
142 until the 1000th cycle, and for every 100th cycle thereafter. Maximum and minimum force and
143 displacement were captured for every cycle. The system's compliance was measured using a
144 custom aluminum part held between the upper and lower clamps that was loaded at $100 \text{ N} \cdot \text{s}^{-1}$ to
145 2000 N. Displacements of the cyclic loading tests were then corrected by subtracting the system's
146 compliance.

147
148 The first ramped loading test was performed prior to the cyclic testing protocol and consisted of a
149 preload test of 5 cycles with a 1 Hz sinusoidal waveform between 0 and 2 MPa, followed by a
150 ramped load from 0 MPa to the tendon-specific peak stress at a rate of $100 \text{ N} \cdot \text{s}^{-1}$. During the

151 ramped load, an AVE2 non-contacting video extensometer (Instron, MS, USA) captured images
152 of the anterior patellar tendon surface at a rate of 50 Hz. The images were 542 mm × 80 mm in
153 size and had a resolution of 0.26 mm/pixel. Subsequent ramped loading tests were performed after
154 stiffness (calculated between 30% and 75% of the peak stress of the loading curve for each cycle)
155 decreased by 10%, 20%, and 30% of the maximum cyclic stiffness. In these instances, the cyclic
156 test was paused, the ramped loading test was completed, and the cyclic test was restarted
157 immediately.

158

159 *Cyclic Test Data Processing*

160 Force data were divided by cross sectional area to calculate stress, and displacement data were
161 converted to nominal strain by dividing displacement (corrected for compliance) by the sample
162 gauge length. Cyclic stress-strain curves were created for every 10% of fatigue life (N_f). The 50th
163 cycle was used for 0% N_f as this allowed the patella and tibia to seat properly within the clamps.
164 Young's modulus was calculated at each percentage of N_f as the slope between 50% and 100% of
165 peak force. Cyclic energy dissipation, calculated as the difference between the area under the
166 loading and unloading stress-strain curves, was numerically computed using the trapezoidal
167 method. A damage parameter was also used to calculate the damage rate per cycle (Cotton et al.,
168 2005):

$$169 \quad D_i = 1 - \frac{E_i}{E_0} \quad (1)$$

170 where E_i is the Young's modulus for a given cycle and E_0 is the initial Young's modulus. Damage
171 rate was calculated as the slope of the damage versus loading cycles curve between 30% and 70%
172 of N_f , as this corresponded to the linear portion of the curve (Figure 5f). Creep rate was calculated

173 as the slope of the peak cyclic strain versus loading cycles, also between 30% and 70% of N_f
174 (Figure 5e).

175

176 *DIC Processing*

177 Commercially available software (DIC Replay; Instron, MS, USA) was used to calculate axial
178 strains from the video extensometer images. Briefly, a region of interest containing the patellar
179 and tibial entheses was created around the tendon, and a 31×31 pixel subset size was used to
180 calculate the displacement of the speckle pattern. The software automatically overlapped
181 neighboring subsets to create a displacement grid spacing equal to one quarter of the subset size.
182 An axial strain colourmap was generated automatically from the displacement grid, which was
183 filtered using a 9×9 pixel filter. Custom Matlab code (version R2017b; MathWorks, MA, USA)
184 was used to extract axial strain at peak stress for all pixels within the colourmap based on their
185 greyscale value. Initial median and 99th percentile axial strains were calculated using the strain
186 distributions from the ramped loading tests. Using the DIC software, a virtual extensometer was
187 placed at the patellar and tibial entheses to obtain an overall measure of initial peak tendon strain
188 (referred to as initial nominal strain).

189

190 *Statistical Analysis*

191 Repeated measures ANOVAs were used to compare Young's modulus and cyclic energy
192 dissipation for every 10% of N_f . A Huynh-Feldt correction was used if the assumption of sphericity
193 was violated, and Bonferroni post hoc tests were performed to examine pairwise differences when
194 a significant main effect was observed. Stress, initial nominal strain, initial median strain, initial
195 99th percentile strain, creep rate, and damage rate were plotted against N_f to assess the relationship

196 between these variables and the fatigue behaviour of the patellar tendon. All statistical analyses
197 were performed using SPSS (version 26; IBM, NY, USA) with a criterion alpha-level of 0.05.

198

199 **Results**

200 A total of 11 samples failed by fracture of the patella or tibia, and it was interesting to note these
201 samples came from donors that weighed less and had lower bone mineral content, as was later
202 confirmed using a quantitative CT mineral analysis of the proximal tibia (Edwards et al., 2013).
203 (Table 1). Of the 19 samples that were successfully tested, 5 had to be re-clamped because some
204 slippage of the bony endpoints in the clamps was observed. These samples were excluded from
205 the analysis of creep rate and damage rate. The exact failure location in the successful samples
206 could not be identified, as fascicles began to progressively fail at an increasing rate until the tendon
207 could not sustain the applied force and ruptured completely. However, we observed that the
208 fascicles tended to rupture near both the patellar and tibial entheses.

209

210 A significant effect of cyclic loading was observed for Young's modulus ($F(2.47, 1) = 101.86, p$
211 < 0.001 ; see Figure 3). Post hoc tests revealed that Young's modulus increased significantly
212 between 0% and 10% N_f ($p = 0.010$), followed by significant decreases from 40% to 50% N_f ($p =$
213 0.029), 70% to 80% N_f ($p < 0.001$), 80% to 90% N_f ($p < 0.001$) and 90% N_f to failure ($p < 0.001$).

214 A significant but less pronounced effect of cyclic loading was also observed for cyclic energy
215 dissipation ($F(1.03,1) = 11.914, p = 0.003$; see Figure 3). Post hoc tests demonstrated significant
216 increases in energy dissipation between 0% and 10% N_f ($p = 0.020$) and between 80% and 90%
217 N_f ($p = 0.022$).

218

219 Relationships between N_f and the outcome variables of stress, strain, creep rate and damage rate
220 were quantified using power law relationships (Figure 5), as these generally displayed better fits
221 than exponential relationships. Peak stress explained 65% of the variance in N_f ($p < 0.001$), and
222 both initial peak nominal strain and initial median strain displayed significant relationships with
223 N_f ($r^2 = 0.57$ and $r^2 = 0.60$, respectively, $p < 0.001$ for both). However, initial 99th percentile strain
224 did not correlate well with N_f ($r^2 = 0.07$, $p = 0.265$). Creep rate had the strongest relationship with
225 N_f , explaining 96% of the variance in N_f ($p < 0.001$), followed by damage rate, which explained
226 86% of the variance in N_f ($p < 0.001$).

227
228 DIC strain distributions visualized at 90%, 80% and 70% of the maximum cyclic stiffness
229 consistently showed increased axial strains near the tendon entheses. Strain concentrations were
230 also present within the tendon midsubstance and tended to increase in size and magnitude
231 throughout the test (see Figure 6).

232

233 **Discussion**

234 The purpose of this study was to quantify the fatigue behaviour and associated material property
235 degradation of human patellar tendon. We observed significant decreases in Young's modulus and
236 increases in cyclic energy dissipation over the course of the cyclic test. Fatigue life was well
237 described by power law relationships for initial nominal strain, initial median strain, and peak
238 stress, but creep rate and damage rate were by far the best predictors. Strain concentrations,
239 calculated using DIC, increased in both size and magnitude over the course of cyclic testing. These
240 data provide important information for our understanding of patellar tendon overuse injuries.

241

242 We observed an increase in Young's modulus at the beginning of cyclic testing, followed by a
243 steady decrease between 30% and 70% N_f , with a sharper decrease from 70% N_f until failure.
244 Expectedly, an opposite trend was observed for cyclic energy dissipation, which initially decreased
245 from 0-10% N_f , and then stabilized until 80% N_f when it sharply increased until failure. These
246 findings indicate that in a fatigue loading scenario, the patellar tendon may be able to sustain a
247 threshold level of energy dissipation until failure is imminent, while tendon stiffness continues to
248 decrease until failure. The observed reductions in Young's modulus and increases in energy
249 dissipation with cyclic loading may be attributed to microdamage accumulation (Fung et al., 2010,
250 2009), which originates from the denaturation of collagen molecules (Zitnay et al., 2020).

251
252 Microdamage accumulation in tendon is indeed strain controlled (Fung et al., 2010; Schechtman
253 and Bader, 1997; Wren et al., 2001; Zitnay et al., 2020), therefore we expected measures of initial
254 strain to be highly correlated with fatigue-life measurements. We discovered that the amount of
255 variance described by the relationship between peak initial strain and the fatigue life of the patellar
256 tendon ($r^2 = 57\%$) was similar to that of the Achilles tendon ($r^2 = 59\%$) from the work of Wren et
257 al. (2003). However, the slope of our power law fit was 4.3, which is considerably less than the
258 slope of 9.3 for the Achilles tendon (Figure 7). While it is tempting to suggest that the fatigue life
259 of Achilles tendon is more sensitive to changes in initial strain than the patellar tendon, it is
260 important to note that fatigue-life measurements from Wren et al., (2003) corresponded to low
261 cycle fatigue (between 10^0 - 10^4 cycles) and displayed more scatter than the patellar tendon results
262 presented herein. Combining the Achilles and patellar tendon data together increased the amount
263 of explained variance substantially ($r^2 = 78\%$, Figure 7), and visual inspection supports the notion
264 that these two tendons illustrate similar fatigue behaviour.

265

266 Human tendon has been observed to rupture at a relatively homogenous peak strain magnitude
267 between 13% and 20% in both monotonic and cyclic testing scenarios (Butler et al., 1986; Cooper
268 et al., 1993; Hashemi et al., 2005; Johnson et al., 1994; Schechtman and Bader, 1997; Wren et al.,
269 2001). Thus, one would expect measures that account for the cyclic- or time-dependent increases
270 in peak strain to be highly correlated with fatigue-life measurements. In this study, the creep rate
271 explained 96% of the variance in fatigue life, which interestingly, is identical to the amount of
272 variance explained by creep rate for human cortical bone samples loaded in cyclic uniaxial tension
273 (Cotton et al., 2005). These findings have important implications for the prediction of overuse
274 injuries in biological tissues. For tendon, if changes to peak cyclic strain over time could be tracked
275 using non-invasive measures (through ultrasound or some other means), this may prove much
276 more valuable than relying on initial estimates of peak stress or strain obtained through
277 biomechanical analysis, for example (Firminger et al., 2020).

278

279 As expected, DIC surface strains increased as the tendon accumulated fatigue damage. We also
280 observed localized areas of increased strain (Figure 6) which did not extend the entire length of
281 the tendon, supporting the notion that load was likely being delivered via shear stress across
282 discontinuous fibrils (Svensson et al., 2013; Szczesny et al., 2015; Szczesny and Elliott, 2014).
283 The heterogeneity of strains across the tendon surface indicates that fatigue damage did not accrue
284 equally across the entire tendon. Rather, weaker areas illustrated increases in strain magnitude and
285 strain concentration size as the tendon stiffness degraded.

286

287 Patellar tendinopathy is considered to be an “insertional tendinopathy” as the symptomatic region
288 is generally located at or near the patellar enthesis. Interestingly, most samples in our study failed
289 near the patellar and tibial entheses. While these observations may be associated with anterior
290 surface strain concentrations which were commonly observed near the entheses, previous research
291 has speculated that patellar tendinopathy may occur due to shear stresses caused by disparities in
292 strain magnitude between the anterior and posterior surfaces near the patellar enthesis (Maganaris
293 et al., 2004). As we were only able to quantify anterior surface strains, future work examining
294 posterior surface strains using a similar protocol may be able to further elucidate the mechanisms
295 of failure near the patellar enthesis.

296

297 Despite decreasing the cross-sectional area of the patellar tendon to avoid specimen pull-out or
298 bone fracture prior to tendon failure, 11 samples fractured pre-emptively at the patella or tibia.
299 Though not an *a priori* objective of this study, we were able to determine that these samples had
300 lower bone mass by analyzing CT images that were originally captured to remove potential
301 specimens with obvious joint pathology. Because the samples that successfully completed the
302 cyclic tests had higher bone mass metrics, we postulated that bone and tendon quality may be
303 correlated in some way. Thus, in a post hoc analysis, we plotted integral measures of bone mineral
304 content (BMC), bone volume (BV), and bone mineral density (BMD) against tendon fatigue life
305 to see whether any relationships existed. None of the aforementioned measures of integral bone
306 mass correlated with fatigue life (BMC, $r^2 = 0.002$; BV, $r^2 = 0.082$; BMD, $r^2 = 0.018$), suggesting
307 that measure of bone mass are not strong surrogate markers of tendon quality.

308

309 The findings from this study are limited to *in vitro* mechanical fatigue tests of cadaveric specimens.
310 The damage state of tendon *in vivo* will, of course, depend on biological repair and adaptation
311 processes, although it is worth noting that tendon may have limited ability for repair (Heinemeier
312 and Kjaer, 2011; Langberg et al., 1999; Miller et al., 2005). Another important limitation of this
313 study is that we were unable to test the full cross-sectional area of the patellar tendon as some of
314 the patella and tibia specimens were likely too weak to withstand the necessary forces required to
315 hold them in the clamp. We acknowledge that removing the medial and lateral portions of the
316 tendon may have altered the tendon's native fatigue properties. However, we were careful to not
317 damage the remaining tendon fascicles and kept a large enough amount of tendon to consider the
318 structure as a subset of the original structure.

319

320 **Conclusions**

321 Patellar tendon samples illustrated decreased stiffness and increased energy dissipation over the
322 course of cyclic testing. Using DIC, we observed the existence of weaker localized strains in areas
323 that grew in size and magnitude as stiffness degraded, illustrating that fatigue damage did not
324 accumulate equally across the entire tendon surface. The fatigue behaviour was well defined by
325 initial nominal strain and initial median strain, further illustrating the strain-dependence of tendon
326 failure in cyclic loading. The excellent correlation between both creep rate and damage rate with
327 fatigue life suggests that non-invasive measurements that track continuous changes in tendon strain
328 may prove effective for preventing overuse injuries of the patellar tendon.

329

330 **Conflicts of interest**

331 The authors have no competing or financial interests to disclose.

332

333 **Funding**

334 Research funding was provided in part by the Natural Sciences and Engineering Research Council
335 of Canada (NSERC; RGPIN 01029-2015, CGSD3 504212-2017), Alberta Innovates, and the
336 NBA/GE Healthcare Orthopedics and Sports Medicine Collaboration. Research infrastructure was
337 funded by a Canadian Foundation for Innovation John R. Evans Leaders Fund (Project #37134).

338

339 **References**

- 340 Arya, S., Kulig, K., 2010. Tendinopathy alters mechanical and material properties of the Achilles
341 tendon. *J. Appl. Physiol.* 108, 670–5. doi:10.1152/jappphysiol.00259.2009
- 342 Butler, D., Kay, M., Stouffer, D., 1986. Comparison of Material Properties in Fascicle-Bone
343 Units From Human Patellar Tendon and Knee Ligaments. *J. Biomech.* 19, 425–432.
344 doi:[https://doi.org/10.1016/0021-9290\(86\)90019-9](https://doi.org/10.1016/0021-9290(86)90019-9)
- 345 Cook, J.L., Khan, K.M., Harcourt, P.R., Grant, M., Young, D.A., Bonar, S.F., 1997. A cross
346 sectional study of 100 athletes with jumper’s knee managed conservatively and surgically.
347 The Victorian Institute of Sport Tendon Study Group. *Br. J. Sports Med.* 31, 332–336.
348 doi:10.1136/bjism.31.4.332
- 349 Cooper, D., Deng, X.H., Burstein, A.L., Warren, R.F., 1993. The strength of the central third
350 tendon graft. *Am. J. Sports Med.* 21, 818–824.
- 351 Cotton, J.R., Winwood, K., Zioupos, P., Taylor, M., 2005. Damage rate is a predictor of fatigue
352 life and creep strain rate in tensile fatigue of human cortical bone samples. *J. Biomech. Eng.*
353 127, 213–219. doi:10.1115/1.1865188
- 354 Edwards, W.B., Schnitzer, T.J., Troy, K.L., 2013. Torsional stiffness and strength of the
355 proximal tibia are better predicted by finite element models than DXA or QCT. *J. Biomech.*
356 46, 1655–1662. doi:10.1016/j.jbiomech.2013.04.016
- 357 Ferretti, A., Puddu, G., Mariani, P.P., Neri, M., 1984. Jumper’s knee: an epidemiological study
358 of volleyball players. *Phys. Sportsmed.* 12, 97–106. doi:10.1080/00913847.1984.11701970
- 359 Firminger, C.R., Asmussen, M.J., Cigoja, S., Fletcher, J.R., Nigg, B.M., Edwards, W.B., 2020.
360 Cumulative Metrics of Tendon Load and Damage Vary Discordantly with Running Speed.
361 *Med. Sci. Sport. Exerc. Publish Ah*, 1549–1556. doi:10.1249/mss.0000000000002287

362 Firminger, C.R., Bruce, O.L., Wannop, J.W., Stefanyshyn, D.J., Edwards, W.B., 2019. Effect of
363 Shoe and Surface Stiffness on Lower Limb Tendon Strain in Jumping, *Medicine & Science*
364 *in Sports & Exercise*. doi:10.1249/MSS.0000000000002004

365 Firminger, C.R., Edwards, W.B., 2016. The influence of minimalist footwear and stride length
366 reduction on lower-extremity running mechanics and cumulative loading. *J. Sci. Med. Sport*
367 *19*, 975–979. doi:10.1016/j.jsams.2016.03.003

368 Fung, D.T., Wang, V.M., Andarawis-Puri, N., Basta-Pljakic, J., Li, Y., Laudier, D.M., Sun, H.B.,
369 Jepsen, K.J., Schaffler, M.B., Flatow, E.L., 2010. Early response to tendon fatigue damage
370 accumulation in a novel in vivo model. *J. Biomech.* *43*, 274–279.
371 doi:10.1016/j.jbiomech.2009.08.039

372 Fung, D.T., Wang, V.M., Laudier, D.M., Shine, J.H., Basta-Pljakic, J., Jepsen, K.J., Schaffler,
373 M.B., Flatow, E.L., 2009. Subrupture tendon fatigue damage. *J. Orthop. Res.* *27*, 264–273.
374 doi:10.1002/jor.20722

375 Gibbon, W.W., Cooper, J.R., Radcliffe, G.S., 1999. Sonographic incidence of tendon microtears
376 in athletes with chronic Achilles tendinosis. *Br. J. Sports Med.* *33*, 129–130.
377 doi:10.1136/bjism.33.2.129

378 Hashemi, J., Chandrashekar, N., Slauterbeck, J., 2005. The mechanical properties of the human
379 patellar tendon are correlated to its mass density and are independent of sex. *Clin. Biomech.*
380 *20*, 645–652. doi:10.1016/j.clinbiomech.2005.02.008

381 Heinemeier, K.M., Kjaer, M., 2011. In vivo investigation of tendon responses to mechanical
382 loading. *J. Musculoskelet. Neuronal Interact.* *11*, 115–123.

383 Johnson, G.A., Tramaglino, D.M., Levine, R.E., Ohno, K., Choi, N.Y., Woo, S.L.Y., 1994.
384 Tensile and viscoelastic properties of human patellar tendon. *J. Orthop. Res.* *12*, 796–803.

385 doi:10.1002/jor.1100120607

386 Kannus, P., Jozsa, L., 1991. Histopathological changes preceding spontaneous rupture of a
387 tendon. *J. Bone Jt. Surg.* 73-A, 1507–1525.

388 Ker, R.F., Wang, X.T., Pike, A.V.L., 2000. Fatigue quality of mammalian tendons. *J. Exp. Biol.*
389 203, 1317–27. doi:10.2165/00007256-199520020-00003

390 Kettunen, J.A., Kvist, M., Alanen, E., Kujala, U.M., 2002. Long-Term Prognosis for Jumper’s
391 Knee in Male Athletes. *Am. J. Sports Med.* 30, 689–692.
392 doi:10.1177/03635465020300051001

393 Langberg, H., Skovgaard, D., Petersen, L.J., Bülow, J., Kjaer, M., 1999. Type I collagen
394 synthesis and degradation in peritendinous tissue after exercises determined by microdialysis
395 in humans. *J. Physiol.* 521, 299–306.

396 Lee, A.H., Elliott, D.M., 2019. Multi-Scale Loading and Damage Mechanisms of Plantaris and
397 Rat Tail Tendons. *J. Orthop. Res.* 37, 1827–1837. doi:10.1002/jor.24309

398 Lee, S.J., Hidler, J., 2008. Biomechanics of overground vs. treadmill walking in healthy
399 individuals. *J. Appl. Physiol.* 104, 747–755. doi:10.1152/jappphysiol.01380.2006

400 Lian, O.B., 2005. Prevalence of Jumper’s Knee Among Elite Athletes From Different Sports: A
401 Cross-sectional Study. *Am. J. Sports Med.* 33, 561–567. doi:10.1177/0363546504270454

402 Luyckx, T., Verstraete, M., De Roo, K., De Waele, W., Bellemans, J., Victor, J., 2014. Digital
403 image correlation as a tool for three-dimensional strain analysis in human tendon tissue. *J.*
404 *Exp. Orthop.* 1, 7. doi:10.1186/s40634-014-0007-8

405 Maganaris, C.N., Narici, M. V, Almekinders, L.C., Maffulli, N., 2004. Biomechanics and
406 Pathophysiology of Overuse Tendon Injuries Ideas on Insertional Tendinopathy 34, 1005–
407 1017.

408 Malliaras, P., Cook, J., Purdam, C., Rio, E., 2015. Patellar tendinopathy: Clinical diagnosis, load
409 management, and advice for challenging case presentations. *J. Orthop. Sports Phys. Ther.*
410 45, 887–898. doi:10.2519/jospt.2015.5987

411 Millar, N.L., Murrell, G.A.C., McInnes, I.B., 2017. Inflammatory mechanisms in tendinopathy -
412 towards translation. *Nat. Rev. Rheumatol.* 13, 110–122. doi:10.1038/nrrheum.2016.213

413 Miller, B.F., Olesen, J.L., Hansen, M., Døssing, S., Cramer, R.M., Welling, R.J., Langberg, H.,
414 Flyvbjerg, A., Kjaer, M., Babraj, J.A., Smith, K., Rennie, M.J., 2005. Coordinated collagen
415 and muscle protein synthesis in human patella tendon and quadriceps muscle after exercise.
416 *J. Physiol.* 567, 1021–1033. doi:10.1113/jphysiol.2005.093690

417 Nakama, L.H., King, K.B., Abrahamsson, S., Rempel, D.M., 2005. Evidence of tendon
418 microtears due to cyclical loading in an in vivo tendinopathy model. *J. Orthop. Res.* 23,
419 1199–1205. doi:10.1016/j.orthres.2005.03.006

420 Noyes, F.R., Butler, D.L., Grood, E.S., Zernicke, R.F., Hefzy, M.S., 1984. Biomechanical
421 Analysis of Human Knee Ligament Grafts used in Knee Ligament Repairs and
422 Reconstructions. *J Bone Jt. Surg AM* 66, 344–352.

423 Palanca, M., Tozzi, G., Cristofolini, L., 2016. The use of digital image correlation in the
424 biomechanical area: A review. *Int. Biomech.* 3, 1–21. doi:10.1080/23335432.2015.1117395

425 Pike, A.V.L., Ker, R.F., Alexander, R.M., 2000. The development of fatigue quality in high- and
426 low-stressed tendons of sheep (*Ovis aries*). *J. Exp. Biol.* 203, 2187–2193.

427 Schechtman, H., Bader, D.L., 2002. Fatigue damage of human tendons. *J. Biomech.* 35, 347–
428 353.

429 Schechtman, H., Bader, D.L., 1997. In vitro fatigue of human tendons. *J. Biomech.* 30, 829–835.

430 Schmid, M., Hodler, J., Cathrein, P., Duewell, S., Jacob, H., Romero, J., 2002. Is impingement

431 the cause of jumper's knee? Dynamic and static magnetic resonance imaging of patellar
432 tendinitis in an open-configuration system. *Am. J. Sports Med.* 30, 388–395.
433 doi:10.1177/03635465020300031401

434 Svensson, R.B., Mulder, H., Kovanen, V., Magnusson, S.P., 2013. Fracture mechanics of
435 collagen fibrils: Influence of natural cross-links. *Biophys. J.* 104, 2476–2484.
436 doi:10.1016/j.bpj.2013.04.033

437 Szczesny, S.E., Aeppli, C., David, A., Mauck, R.L., 2018. Fatigue loading of tendon results in
438 collagen kinking and denaturation but does not change local tissue mechanics. *J. Biomech.*
439 71, 251–256. doi:10.1016/j.jbiomech.2018.02.014

440 Szczesny, S.E., Caplan, J.L., Pedersen, P., Elliott, D.M., 2015. Quantification of interfibrillar
441 shear stress in aligned soft collagenous tissues via notch tension testing. *Sci. Rep.* 5, 1–9.
442 doi:10.1038/srep14649

443 Szczesny, S.E., Elliott, D.M., 2014. Interfibrillar shear stress is the loading mechanism of
444 collagen fibrils in tendon. *Acta Biomater.* 10, 2582–2590. doi:10.1016/j.actbio.2014.01.032

445 Tallon, C., Maffulli, N., Ewen, S.W.B., 2001. Ruptured Achilles tendons are significantly more
446 degenerated than tendinopathic tendons. *Med. Sci. Sports Exerc.* 33, 1983–1990.
447 doi:10.1097/00005768-200112000-00002

448 Thorpe, C.T., Chaudhry, S., Lei, I.I., Varone, A., Riley, G.P., Birch, H.L., Clegg, P.D., Screen,
449 H.R.C., 2015. Tendon overload results in alterations in cell shape and increased markers of
450 inflammation and matrix degradation. *Scand. J. Med. Sci. Sport.* 25, e381–e391.
451 doi:10.1111/sms.12333

452 Wang, X.T., Ker, R.F., Alexander, R.M., 1995. Fatigue rupture of wallaby tail tendons. *J. Exp.*
453 *Biol.* 198, 847–852.

454 Wren, T.A.L., Lindsey, D.P., Beaupre, G.S., Carter, D.R., 2003. Effects of creep and cyclic
455 loading on the mechanical properties and failure of human Achilles tendons. *Ann. Biomed.*
456 *Eng.* 31, 710–717. doi:10.1114/1.1569267

457 Wren, T.A.L., Yerby, S.A., Beaupré, G.S., Carter, D.R., 2001. Mechanical properties of the
458 human achilles tendon. *Clin. Biomech.* 16, 245–251. doi:10.1016/S0268-0033(00)00089-9

459 Zitnay, J.L., Jung, G.S., Lin, A.H., Qin, Z., Li, Y., Yu, S.M., Buehler, M.J., Weiss, J.A., 2020.
460 Accumulation of collagen molecular unfolding is the mechanism of cyclic fatigue damage
461 and failure in collagenous tissues. *Sci. Adv.* 6.

462

Tables

Table 1: Summary of donor information and indices of tibial integral bone mineral

Test Result	Age (years)	Height (cm)	Weight (kg)	BMI (kg/m ²)	Sex (male/female)	BV (cm ³)	BMC (g)	BMD (g/cm ³)
Successful	73.6 (12.7)	172.5 (8.9)	77.3 (15.6)	25.9 (4.4)	14/5	192.41 (46.36)	45.04 (18.14)	0.23 (0.09)
Unsuccessful	80.8 (12.8)	168.3 (9.2)	66.4 (9.0)	24.7 (3.7)	5/6	162.08 (36.21)	28.42 (21.49)	0.16 (0.10)
All	76.2 (14.3)	170.9 (9.1)	73.3 (14.4)	25.5 (4.1)	19/11	181.58 (44.83)	39.10 (20.66)	0.21 (0.10)
p-value	0.187	0.244	0.043	0.458	-	0.086	0.039	0.063

Independent samples t-tests were performed to assess statistical differences between successful and unsuccessful tests.

Figures

Figure 1: a) Schematic of sample preparation; the tibia was transected 40 mm distal to the tibial tuberosity and was machined to fit into the upper clamp. The proximal 10 mm of the tibia was also removed to allow for adequate spacing between the tibia and the lower clamp. b) Experimental setup of the fatigue test. The prepared sample was inverted, with the tibia held in the upper clamp and the patella held in the lower clamp at an angle of 146° to the patellar tendon. Four saline drips kept the sample hydrated during testing.

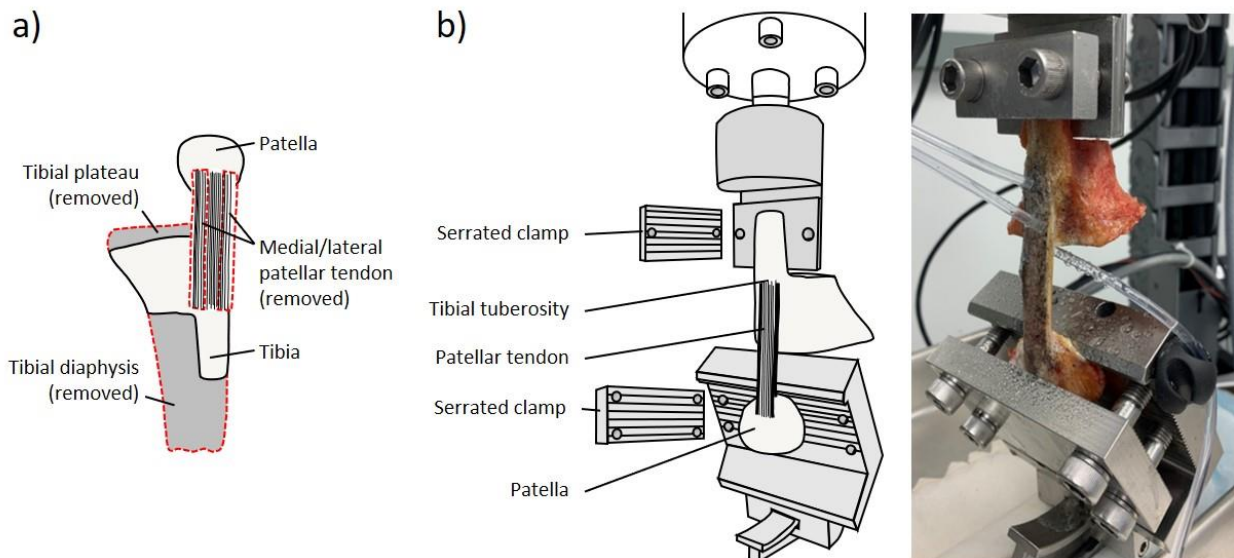


Figure 2: Stiffness profile of a representative sample cyclically loaded between 2 MPa and the test stress at 2 Hz until failure (red X). Ramped tests with DIC were performed prior to cyclic loading, and at 90%, 80% and 70% of maximum cyclic stiffness (k_{max}).

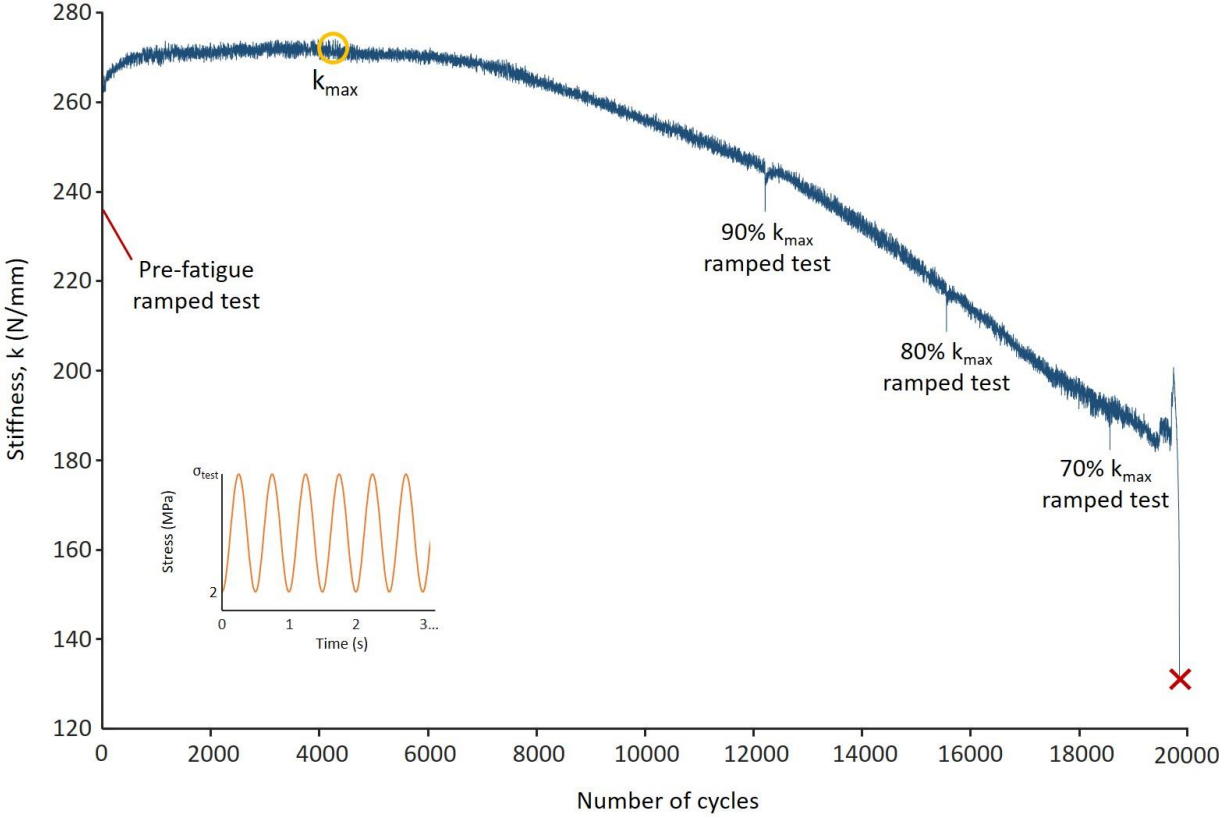


Figure 3: Stress-strain cycles 0-100% fatigue life (N_f) for a representative sample.

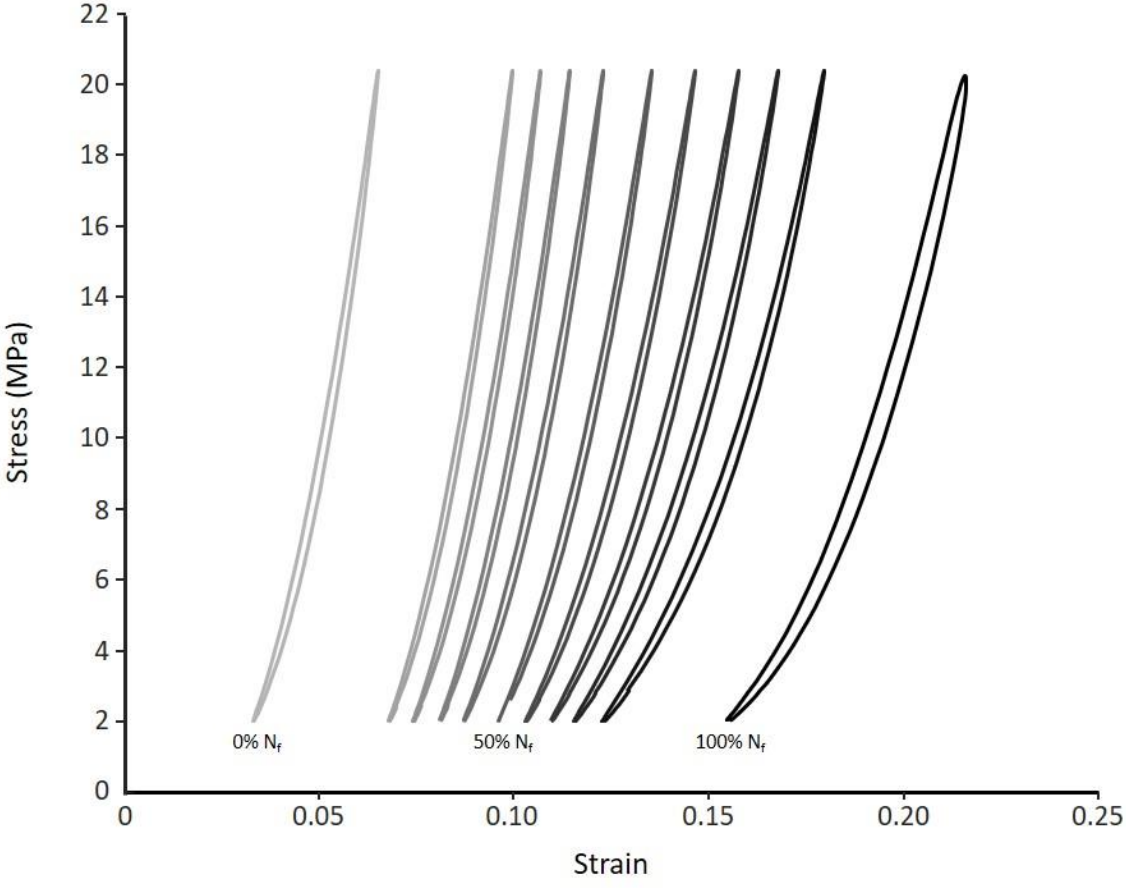


Figure 4: Estimated marginal means \pm standard deviation for: a) Young's modulus and b) energy dissipation of stress-strain cycles from 0-100% fatigue life (N_f). *Significant difference ($p < 0.05$).

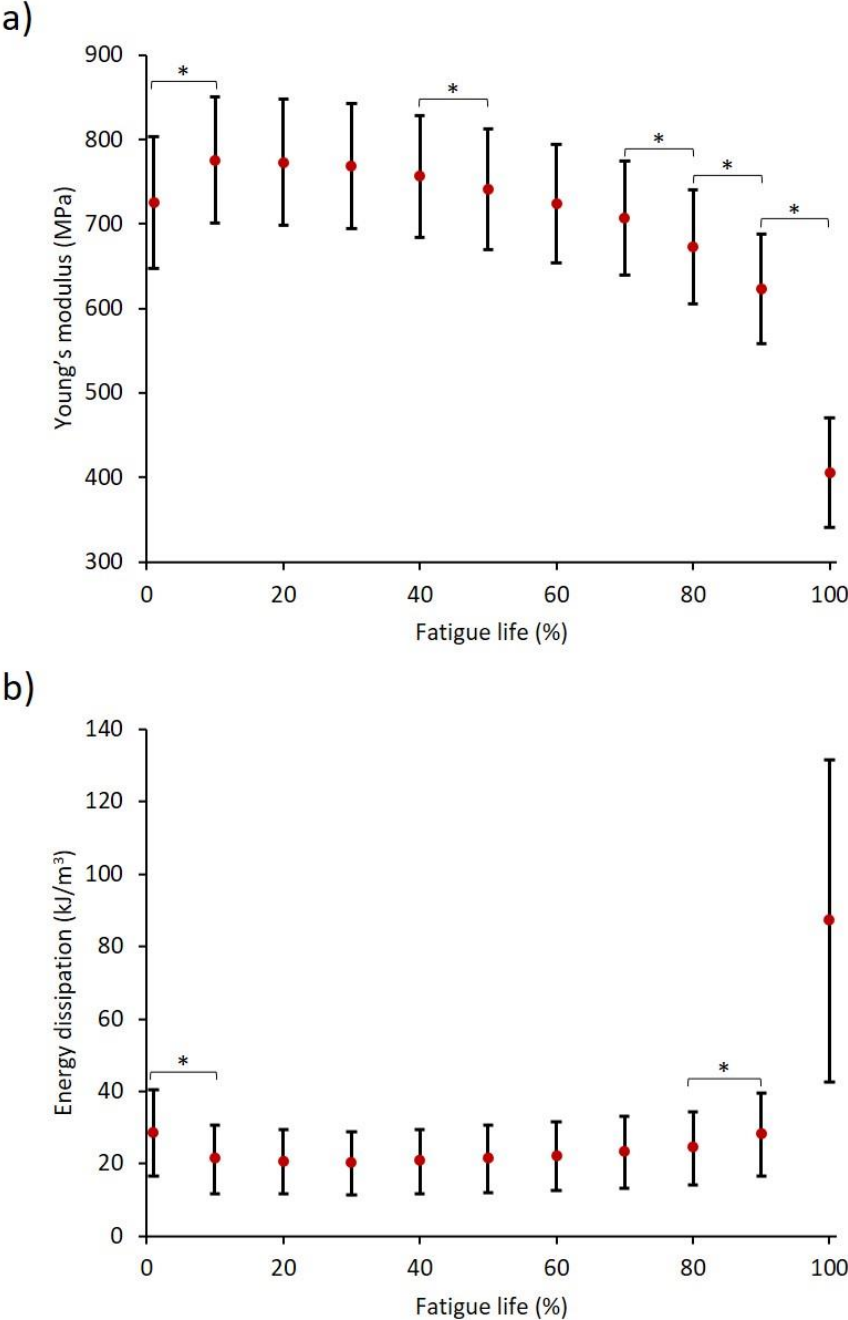


Figure 5: Plots of a) peak stress, b) initial peak nominal strain, c) initial median strain, d) initial 99th percentile strain, e) creep rate, and f) damage rate, versus fatigue life. All plots have been fitted with a power law curve. The insets in e) and f) illustrate how creep rate and damage rate were calculated using a representative sample.

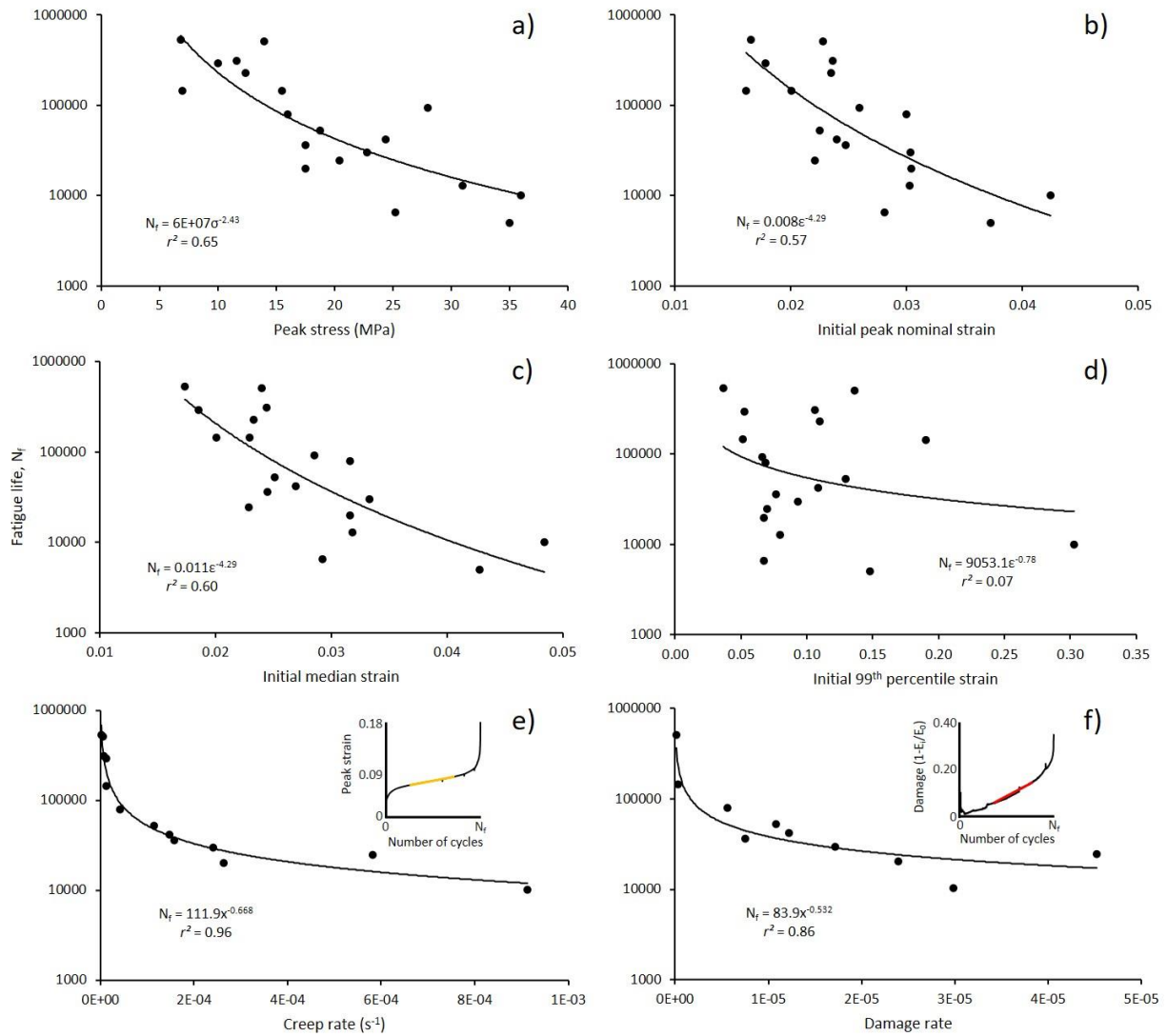


Figure 6: Axial surface strains of a representative sample: a) prior to the fatigue test; b) at 90% of the maximum cyclic stiffness; c) at 80% of the maximum cyclic stiffness; d) at 70% of the maximum cyclic stiffness.

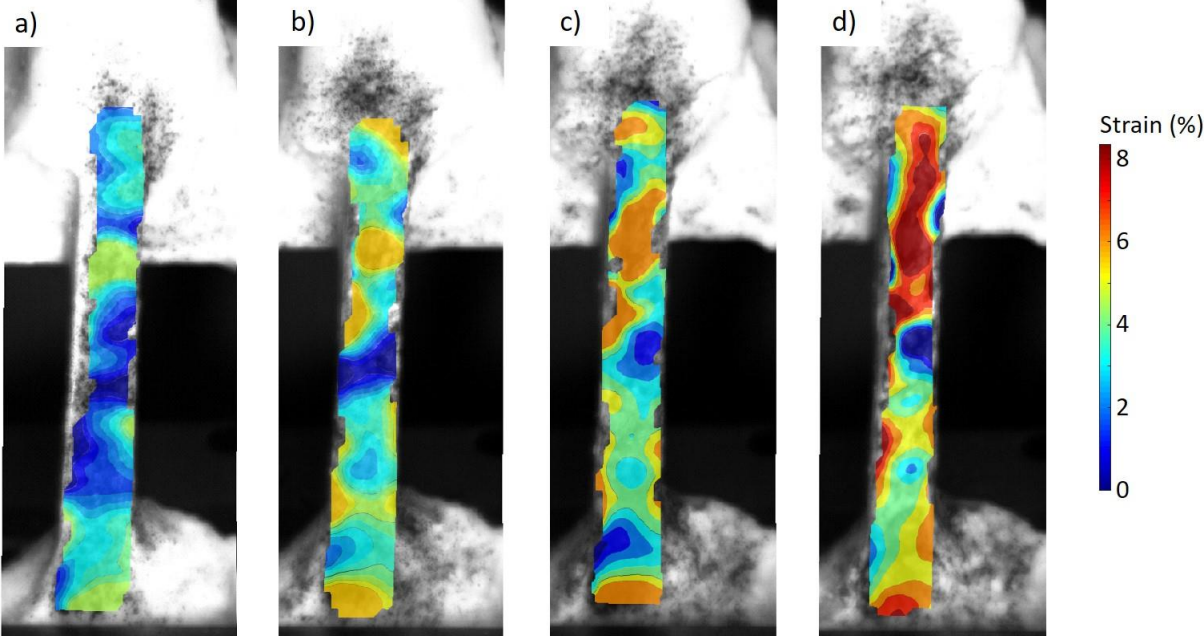


Figure 7: Initial nominal strain versus fatigue life for the patellar and Achilles tendons (red and blue points, respectively). Achilles tendon data were obtained from Wren et al (2003). Individual power law curves fitted to patellar ($N_{f,PT}$) and Achilles ($N_{f,AT}$) tendon data illustrated similar amounts of explained variance. Pooling the patellar and Achilles tendon data ($N_{f,Both}$) explained a larger amount of variance than the individual curves.

